See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/273206722

# Evidence of sewage-driven eutrophication and harmful algal blooms in Florida's Indian River Lagoon

Article in Harmful Algae · March 2015

DOI: 10.1016/j.hal.2015.01.004

citation 40	IS	READS 839	
4 autho	ors, including:		
	Brian E. Lapointe Florida Atlantic University 102 PUBLICATIONS 5,521 CITATIONS SEE PROFILE	Laura W. HerrenFlorida Atlantic University7 PUBLICATIONS87 CITATIONSEE PROFILE	
	Margaret Vogel Florida State University <b>3</b> PUBLICATIONS <b>57</b> CITATIONS SEE PROFILE		
Some o	of the authors of this publication are also	working on these related projects:	

Project

Great White Heron National Wildlife Refuge View project



Negril Marine Park View project

All content following this page was uploaded by Brian E. Lapointe on 12 February 2018.

The user has requested enhancement of the downloaded file.

Contents lists available at ScienceDirect

# Harmful Algae

journal homepage: www.elsevier.com/locate/hal

# Evidence of sewage-driven eutrophication and harmful algal blooms in Florida's Indian River Lagoon



HARMEU

## Brian E. Lapointe<sup>\*</sup>, Laura W. Herren, David D. Debortoli, Margaret A. Vogel

Harbor Branch Oceanographic Institute at Florida Atlantic University, Harmful Algal Bloom Program, 5600 US 1 North, Fort Pierce, FL 34946, USA

#### ARTICLE INFO

## ABSTRACT

Article history: Received 17 October 2014 Received in revised form 27 January 2015 Accepted 28 January 2015

Keywords: HAB Nitrogen Phosphorus Indian River Lagoon Eutrophication Stable isotopes coastal waters worldwide. In 2011–2012, 20 sites evenly distributed throughout the 251-km long Indian River Lagoon (IRL) were assessed during three sampling events for dissolved nutrients (DIN, SRP, TDN, TDP) and chlorophyll a. Benthic macroalgae were also analyzed for  $\delta^{13}$ C,  $\delta^{15}$ N, and C:N:P contents to identify potential nutrient sources and gauge the type and degree of N and P limitation. The mean DIN and SRP concentrations throughout the IRL were high, averaging  $4.24\pm0.45$  and  $0.68\pm0.06\,\mu\text{M},$ respectively, explaining the widespread occurrence of HABs during the study. High TDN concentrations (up to 152  $\mu$ M) and TDN:TDP ratios (>100:1) in the poorly flushed northern IRL, Mosquito Lagoon and Banana River segments reflected the accumulation and cycling of N-rich groundwater inputs that produce Plimitation. These enriched nutrient conditions were associated with unprecedented chlorophyll a concentrations (>100  $\mu$ g/L), dominated by *Resultor* sp. Ø. Moestrup in the Banana River in 2011 and Aureoumbra lagunensis D.A. Stockwell, DeYoe, Hargraves and P.W. Johnson in the Mosquito Lagoon and northern IRL in 2012. C:N, C:P, and N:P ratios in macroalgae averaged 15.9, 698.9, and 40.6, throughout the IRL, respectively; significantly higher C:P and N:P ratios in the northern IRL segments suggested strong Plimitation in these N-enriched waters. Macroalgae  $\delta^{15}$ N values were enriched throughout the IRL (+6.3‰) and similar to values reported for macroalgae from other sewage-polluted coastal waters. Because pointsource sewage inputs to the IRL were largely eliminated through the IRL Act of 1990, these results suggest that non-point source N enrichment from septic tanks (~300,000) represents a significant and largely ignored N-source to the IRL. The high degree of sewage N contamination of the IRL, combined with recent HABs, including toxic ecotypes of the red macroalga Gracilaria tikvahiae McLachlan, seagrass loss, and wildlife mortality, indicates a critical need for improved sewage collection and treatment, including nutrient removal

Nutrient pollution is a primary driver of eutrophication and harmful algal blooms (HABs) in estuaries and

© 2015 Elsevier B.V. All rights reserved.

#### 1. Introduction

Coastal and estuarine ecosystems are among the most productive ecosystems in the world, providing invaluable ecological services to human populations. However, many of these ecosystems are being degraded as a result of expanding human activities such that their ability to sustain future societal needs is now at risk. Humans have greatly increased the concentrations of nitrogen (N) and phosphorus (P) in freshwaters flowing into the coastal zone (Nixon, 1995; Vitousek et al., 1997; MEA, 2005), exacerbating eutrophication and habitat loss (NRC, 2000; Bricker et al., 2007). As a result, nutrient enrichment is now a major agent

\* Corresponding author. Tel.: +1 772 242 2276. E-mail address: blapoin1@fau.edu (B.E. Lapointe).

http://dx.doi.org/10.1016/j.hal.2015.01.004 1568-9883/© 2015 Elsevier B.V. All rights reserved. of global change in coastal waters, linking an array of problems along coastlines, including eutrophication, biodiversity loss, harmful algal blooms (HABs), "dead zones," emerging marine diseases, fish kills, and loss of seagrass and coral reef ecosystems (NRC, 2000; Howarth and Marino, 2006; Rockström et al., 2009).

Located along Florida's east-central coast, the Indian River Lagoon (IRL) is a shallow (mean depth ~0.8 m) and narrow (~3 km wide) bar-built estuary extending 251 km between Jupiter and Ponce inlets (Steward and VanArman, 1987; Fig. 1). Because the IRL comprises a transition zone between temperate and subtropical biomes, the IRL is considered a regional-scale ecotone and one of the most species-diverse estuaries in North America (Swain et al., 1995). The basin includes the Mosquito Lagoon (ML) and Banana River (BR), which are located in the northern regions of the IRL. The climate of the IRL basin is humid subtropical with distinct dry and wet seasons. Rainfall within the basin averages 140–150 cm/yr,



with 62% falling from June through October (IRLNEP, 1996). Freshwater enters the IRL basin from rainfall, surface water runoff, submarine groundwater discharge (SGD), and inflows from tributaries and canals.

The IRL watersheds have experienced dramatic changes in landuse over the past century. Historically, drainage of the IRL basin occurred through slow, meandering streams, creeks, rivers, and wetlands. Since the Drainage Acts of Florida (1916) that permitted the creation of canals to drain uplands for agriculture, reduce flooding, and control mosquitos, the IRL watershed has nearly tripled its size from 231,480 hectares to more than 566,560 hectares (SJRWMD, 2007). These changes greatly altered the hydrology and increased stormwater discharges to the IRL. This is especially evident in the St. Lucie Estuary (SLE; Fig. 1), which now receives periodic excessive freshwater discharges from Lake Okeechobee, especially following hurricanes and tropical storms (Lapointe et al., 2012). Since the drainage of the IRL watersheds, the increased land area and accompanying socio-economic opportunities directly contributed to rapid urbanization and population increases. The population in the IRL region has increased from ~250,000 in 1960 to ~1.7 million today (US Census Bureau, 2014). As a consequence, land use changes in the IRL watershed have been dramatic. In 1920, only 4% of the land was classified as low-density residential, while 95% of land was forest/ grass/pasture and 1% agriculture (Kim et al., 2002). The IRL watershed is now dominated by urban land uses (39%), followed by agriculture (24%), range (20.8%), wetland (12.1%), and natural forest (4.5%; Bricker et al., 2007).

Eutrophication of the IRL resulting from widespread urbanization and population growth has long been a concern of scientists and resource managers. The number of tidal inlets that allow flushing with Atlantic coastal waters is limited and the residence time of water in the northern IRL can be high (>1 yr; Smith, 1993). High residence times combined with rapid population growth on its watersheds, has made the northern IRL especially susceptible to nutrient enrichment, eutrophication, and seagrass die-off (Briel

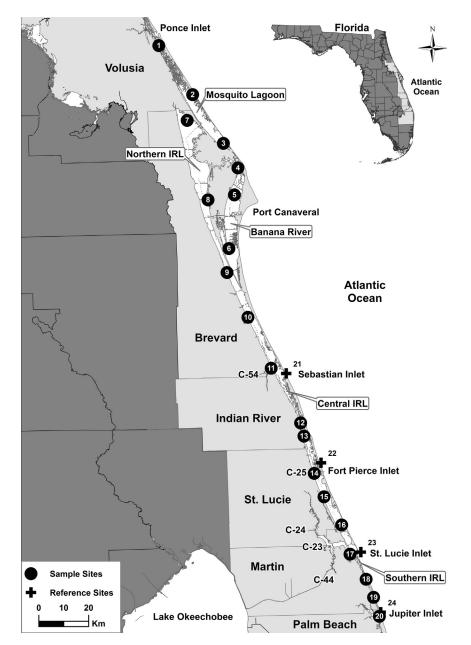


Fig. 1. Map of the Indian River Lagoon showing the five segments, 20 sampling sites, and four reference sites.

et al., 1973; Bricker et al., 2007; FDEP, 2008). Increasing land-based sources of nutrients in surface waters of the IRL primarily originate from stormwater and groundwater within agricultural and urban areas on the watershed. In recent decades, urbanization of the watershed, especially in the northern IRL segments, has increased rapidly with parallel decreases in agriculture (Duncan et al., 2004). In 1990, concerns of increased sewage-driven eutrophication led to the Indian River Lagoon Act of 1990 (IRL Act; Chapter 90-262, Laws of Florida) that required sewage treatment plants to cease discharging into surface waters of the IRL by July 1, 1995. The IRL Act also required municipalities to identify areas where package sewage treatment plants and septic tanks posed threats to the IRL and implement plans to provide centralized sewage treatment to these areas by July 1, 1994. By 1996, point-source sewage discharges (outfalls) into surface waters of the IRL were largely eliminated (IRLNEP, 2008). The Watershed Restoration Act of 1999 (403.067 F.S.) mandated the Florida Department of Environmental Protection (FDEP) to scientifically evaluate the quality of Florida's surface waters and promote the mechanisms necessary to reduce nutrient pollution through the implementation of the federal Total Maximum Daily Load (TMDL) program. The effort of the implementation was mainly geared toward the reduction of nutrient inputs from wastewater treatment plants (WWTPs) and stormwater runoff. With the exception of periodic wet weather discharges, all WWTPs in the IRL region are presently in compliance with current water quality protection codes, rules, and statues (IRLNEP, 2008).

Despite the elimination of point-source sewage inputs to the IRL through the IRL Act, non-point source sewage pollution from septic tanks (on-site sewage treatment and disposal systems, OSTDS) has continued to expand and remains a serious environmental and human health concern (Bicki et al., 1984; Lapointe and Krupa, 1995a,b; Belanger et al., 2007; Lapointe et al., 2012). Federal and state agencies have long recognized the harmful impacts of OSTDS to groundwaters and surface waters in Florida, including the IRL (Bicki et al., 1984; Woodward-Clyde, 1994; IRLNEP, 1996; Kroening, 2007; IRLNEP, 2008). Currently, there are an estimated 300,000 OSTDS installed in the counties adjacent to the IRL, with the majority of these located in Brevard and Volusia counties, which border the poorly-flushed ML, BR, and northern IRL (Fig. 1). Contamination of groundwaters with nutrients derived from non-sewered human wastewater has long been known to contribute to eutrophication of lakes, estuaries, and coastal waters (Brezonik, 1972; Bicki et al., 1984; Lapointe et al., 1990; Valiela et al., 1992; Weiskel and Howes, 1992; NRC, 1993; US EPA, 2002) and could be an important pathway for nutrient pollution of the IRL (Martin et al., 2007). Such groundwater-borne nutrient inputs would negatively impact the seagrass communities and biodiversity of the IRL by fueling harmful algal blooms (HABs), including macroalgae and/or epiphytes (Lapointe et al., 1994; Burkholder et al., 2007) and phytoplankton (Phlips et al., 2011; Lapointe et al., 2012; Gobler et al., 2013). Groundwaters contaminated by OSTDS have impacted a wide variety of other urbanized estuaries and coastal waters, including Bermuda (Lapointe and O'Connell, 1989; McGlathery, 1995), the Florida Keys (Lapointe et al., 1990; Tomasko and Lapointe, 1991; Lapointe et al., 1994), Cape Cod, MA (Valiela et al., 1992; Weiskel and Howes, 1992; Valiela et al., 1997), Outer Banks, North Carolina (Mallin, 2013), Chesapeake Bay (Reay, 2004), and Puerto Rico (Olsen et al., 2010). In addition, bacterial contamination of groundwaters and surface waters has been associated with unsuitable soil characteristics and high densities of OSTDS on the IRL watersheds (Lapointe and Krupa, 1995a,b; Belanger et al., 2007; Lapointe et al., 2012) and is considered a primary source of bacterial contamination associated with immunologic perturbations in populations of bottlenose dolphins, *Tursiops truncatus* Montagu, in the IRL (Schaefer et al., 2009; Bossart et al., 2014). Schaefer et al. (2011) found a positive correlation between the number of IRL bottlenose dolphins colonized by *Escherichia coli* and the number of septic tanks in the area they lived; with the highest number of both being in the northern segments of the IRL. Similarly, the health of juvenile green sea turtles *Chelonia mydas* Linnaeus as measured through blood parameters and the severity and prevalence of fibropa-pillomatosis (caused by an alphaherpesvirus) was significantly poorer in the IRL population than those captured on adjacent nearshore wormrock reefs (Hirama et al., 2014).

Blooms of benthic macroalgae have been symptomatic of Ndriven eutrophication in the shallow waters of the IRL for decades (Lapointe and Ryther, 1979; Benz et al., 1979; Virnstein and Carbonara, 1985; Bricker et al., 2007) and provide effective indicator organisms to assess the relative importance of N sources and gauge the type and degree of N vs. Plimitation. Macroalgae are ideal "bio-observatories" for assessing nutrient availability as they are typically attached to the benthos and integrate nutrient availability over temporal scales of days to weeks (Lapointe, 1985). Measurement of stable carbon and nitrogen isotopes ( $\delta^{13}$ C,  $\delta^{15}$ N) in macroalgal tissue have been widely used to discriminate among natural (upwelling, N-fixation) and anthropogenic (sewage, fertilizers) nutrient sources (Risk et al., 2008). Because natural N-fixation source values are close to 0‰ (Heaton, 1986; France et al., 1998), atmospheric N typically ranges from -3‰ to +1‰ (Paerl and Fogel, 1994) and synthetic fertilizer N ranges from -2% to +2% (Bateman and Kelly, 2007); these N sources are all depleted relative to enriched values of +3% to +19% for human sewage (Heaton, 1986; Costanzo et al., 2001; Table 1). Accordingly, enriched  $\delta^{15}N$  values > +3% have been reported for macroalgae in a wide variety of sewage-polluted coastal waters, including Boston Harbor, MA (France et al., 1998), Childs River, Cape Cod, MA (McClelland and Valiela, 1998), Narrangansett Bay, RI (Thornber et al., 2008), Sarasota Bay, FL (Lapointe, 2013), Moreton Bay, Australia (Costanzo et al., 2001), and nearshore reefs off urban areas of east-central Florida (Barile, 2004), southeast Florida (Lapointe et al., 2005a), southwest Florida (Lapointe and Bedford, 2007), Jamaica (Lapointe et al., 2011), and Tobago (Lapointe et al., 2010; Table 1). In addition, measurement of C:N:P contents of macroalgae provides a measure of nutrient quantity and stoichiometry that is useful in assessing the relative importance of N vs. P-limitation (Atkinson and Smith, 1983; Lapointe et al., 1992). This is particularly appropriate for assessing OSTDS groundwater-borne sewage pollution that can deliver nutrient pollution at high N:P ratios as a result of selective adsorption of Ponto soil particles (Bicki et al., 1984; Lapointe et al., 1990; Weiskel and Howes, 1992).

To assess spatial and temporal variability in N source(s) and overall patterns of eutrophication and nutrient limitation of HABs in the IRL, we performed a comprehensive two-year study throughout the IRL. Previous water quality studies of the IRL have largely focused on the region north of the Ft. Pierce Inlet (Sigua et al., 2000; Steward et al., 2005); in comparison, we chose to perform an IRL-wide study using 20 fixed stations evenly spaced among the five IRL segments (Mosquito Lagoon [ML], Banana River [BR], Northern IRL [NIRL], Central IRL [CIRL], and Southern IRL [SIRL]) extending from Ponce Inlet in Volusia County to Jupiter Inlet in northern Palm Beach County; we also included four reference stations located just outside the IRL on the nearshore sabellarid wormrock reefs (Fig. 1). The study involved sampling the water column for various forms of dissolved nutrients (N and P) and chlorophyll *a*, macroalgae for  $\delta^{13}$ C,  $\delta^{15}$ N, and C:N:P ratios, and seagrass communities for species composition and percent cover to assess nutrient stress in this key management endpoint.

#### Table 1

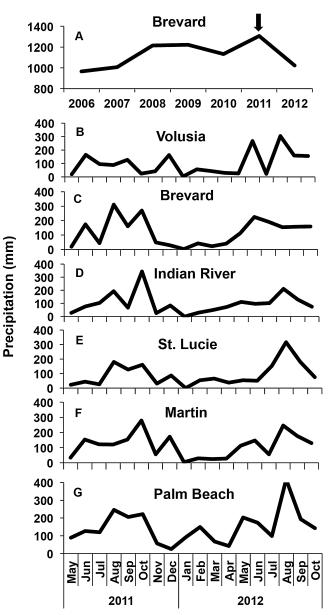
 $\delta^{15}$ N levels reported for macroalgae in both sewage-impacted and relatively unpolluted reference waters worldwide.

Location	Species	Mean $\delta^{15}$ N‰	Reference
Sewage impacted			
Boston Harbor, MA, USA	Ceramium sp.	$X = 6.5 \pm 0.7$ (SD)	France et al. (1998)
	Chondrus crispus		
	Desmarestia sp.		
	Ectocarpus sp.	V 75 45	
Narrangansett Bay, RI, USA	Ulva spp.	X = 7.5 - 15	Thornber et al. (2008)
Childs River, Cape Cod, MA, USA	Cladophora vagabunda Enteromorpha sp.	$5.4 \pm 0.1$ (SE) $8.4 \pm 0.2$ (SE)	McClelland and Valiela (1998)
	Gracilaria tikvahiae	$7.6 \pm 0.4$ (SE)	
Sage Lot Pond, Cape Cod, MA, USA	Cladophora vagabunda	$3.4 \pm 0.1$ (SE)	McClelland and Valiela (1998)
bage lot rona, cape coa, mil, corr	Enteromorpha sp.	$4.9 \pm 0.1$ (SE)	meetenana ana vanera (1999)
	Gracilaria tikvahiae	$5.1 \pm 0.6$ (SE)	
Nearshore reefs, east-central FL, USA	Botryocladia spinulifera	X=8.7-9.9	Barile (2004)
	Bryothamnion triquetrum		
	Caulerpa spp.		
	Chaetomorhpa linum		
	Codium isthmocladum		
	Colpomenia sinuosa		
	Enteromorhpa intestinalis		
	Gracilaria tikvahiae		
	Laurencia poiteaui		
Coastal mode Coutboast EL LICA	Ulva lactuca Caulerpa spp.	$X = 5.52 \pm 0.88$ (SE)	Langinto et al. (2005)
Coastal reefs, Southeast FL, USA	Codium isthmocladum	$X = 5.52 \pm 0.88$ (SE) $X = 6.95 \pm 0.97$ (SE)	Lapointe et al. (2005)
Coastal Reefs, Southwest FL, USA	Agardhiella subulata	$X = 0.93 \pm 0.97$ (SE) $X = 3.89 \pm 0.96 - 5.84 \pm 1.37$ (SD)	Lapointe and Bedford (2007)
coastal Accis, Southwest FE, OSA	Botryocladia occidentalis	$X = 3.85 \pm 0.50 = 3.84 \pm 1.57$ (3D)	Laponite and Dediord (2007)
	Cladophora sp.		
	Eucheuma isiforme var. denudatum		
	Gracilaria spp.		
	Hypnea musciformis		
	Rhodymenia divaricata		
Sarasota Bay, FL, USA	Acanthophora spicifera	$X = 3.76 \pm 0.13$	Lapointe (2013)
	Botryocladia occidentalis		
	Caulerpa spp.		
	Gracilaria spp.		
	Hypnea spp.		
Jrbanized canals, Florida Keys, USA	Chaetomorpha spp.	$X = 4.31 \pm 0.35$ (SE)	Lapointe, unpublished data
	Caulerpa spp.		
Shallow reefs, South Negril, Jamaica	Halimeda opuntia Acanthophora spicifera	$V = 4.70 \pm 0.52$ (SE)	Lapointe et al. (2011)
Shahow reels, South Negrii, Janiaica	Bryothamnion triquetrum	$X = 4.79 \pm 0.53$ (SE)	Lapointe et al. (2011)
	Chaetomorpha gracilis		
	Chaetomorpha linum		
	Cladophora fuliginosa		
	Codium isthmocladum		
	Sargassum spp.		
	Spyridia hypnoides		
Shallow reefs, Buccoo Reef Complex, Tobago	Bryopsis spp.	$X = 6.2 \pm 1.1$ (SD)	Lapointe et al. (2010)
	Caulerpa spp.		
	Dictyota spp.		
	Halimeda opuntia		
	Laurencia poiteaui		
Moreton Bay, Australia	Catenella nipae	2.5-11.3	Costanzo et al. (2001)
	Gracilaria edulis		
Reference sites			
Coral Reefs, Green Turtle Cay, Abacos, Bahamas	Caulerpa verticillata	$X = 1.75 \pm 0.06$ (SD)	Lapointe et al. (2005b)
-	Caulerpa racemosa	$X = 0.78 \pm 0.37$ (SD)	· ·
Coral Reefs, Puerto Rico	Acanthophora spicifera	$X = 0.3 \pm 1.0 \text{ (SD)}$	France et al. (1998)
	Avrainvillea longicaulis		
	Halimeda incrassata		
	Penicillus capitatus		

### 2. Methods

Spatial and temporal variability in water quality throughout the IRL were documented by collection of seawater and macroalgal tissue from the two most abundant species at 20 fixed IRL sampling sites plus four fixed reference sites (REF) on nearshore sabellarid wormrock reefs (Fig. 1) during three sampling events in 2011 and 2012. The 20 IRL sites were grouped by waterbody segment, which

included ML (n = 3), BR (n = 3), NIRL (n = 4), CIRL (n = 5), and SIRL (n = 5). Because the IRL has been increasingly impacted by human activities over the past century, rainfall now plays a significant role in stormwater-driven nutrient pollution and the overall health of the system. Rainfall patterns prior to and during the study showed the transition from the end of a prolonged multi-year drought followed by more typical "wet vs. dry" rainfall seasonality during the two-year study (SJRWMD et al., 2012; Fig. 2A). Accordingly, the



**Fig. 2.** Total annual precipitation in Brevard County near center of superbloom (A; arrow represents year of superbloom; Weather station GHCND: USW00092821) and monthly precipitation in each of the six counties bordering the Indian River Lagoon (B–G) during the 2011–2012 study period (NOAA National Climatic Data Center).

three successive sampling events were classified as Dry 2011 (June 2, 2011–July 13, 2011), Wet 2011 (November 4, 2011–January 11, 2012), and Wet 2012 (August 7, 2012–September 21, 2012; Fig. 2B–G).

#### 2.1. Collection and analysis of water column nutrients and chlorophyll a

Seawater samples were collected in triplicate 0.25 m below the surface into acid-washed 0.5 L high-density polyethylene (HDPE) bottles and covered with ice in a dark cooler until return to the Harbor Branch Oceanographic Institute (HBOI) laboratory for processing. Sampling sites adjacent to canals and tributaries (e.g., 10, 11, 12, 13, 14, 17, and 20 on Fig. 1) were sampled during an ebbing tide. The samples were filtered (0.7  $\mu$ m GF/F filters) and frozen until analysis at the Nutrient Analytical Services

Laboratory, Chesapeake Biological Laboratory, Solomons, MD. The samples were analyzed on a Technicon Auto-Analyzer II (nitrate, TDN, SRP, TDP) or a Technicon TRAACS 800 (ammonium, nitrite). Detection limits were 0.21  $\mu$ M for ammonium, 0.01  $\mu$ M for nitrate and nitrite, 0.02  $\mu$ M for SRP, 2.06  $\mu$ M for TDN, and 0.05  $\mu$ M for TDP. The resulting data were used to characterize ambient dissolved inorganic and total N and P concentrations, DIN:SRP ratios, and TDN:TDP ratios at the IRL sites. Calibrated YSI<sup>TM</sup> Models 63 and 85 hand-held meters were used to determine pH and salinity, conductivity, temperature, and dissolved oxygen, respectively at the time water samples were collected at each site.

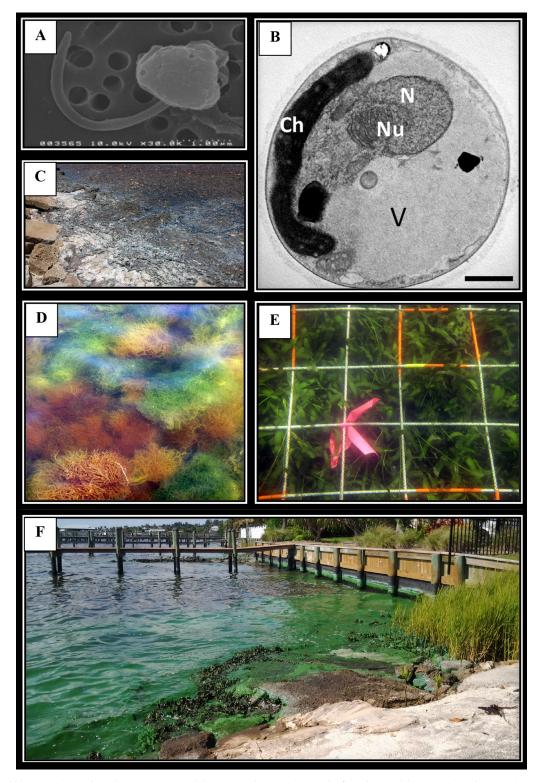
For chlorophyll *a* analysis, GF/F filters were frozen until extracted. To extract, the filters were placed in a 15 mL centrifuge tube with 10 mL of 90% acetone, ground with a Telfon pestle, and allowed to extract for 2–24 h in the dark under refrigeration. After extraction, the samples were removed from refrigeration, warmed to room temperature, and centrifuged at ~2400 rpm for ~10 min. The samples were measured fluorometrically before and after acidification using 5% HCl for chlorophyll *a* and phaeopigment concentrations. Fluorescence measurements were made with a Turner Designs TD700 fluorometer equipped with a daylight white lamp, 340–500 nm excitation filter and >665 nm emission filter or with a Turner Designs Trilogy fluorometer.

#### 2.2. $\delta$ 13C, $\delta$ 15N, and C:N:P analysis of HABs

Blooms of both macroalgae and phytoplankton were sampled to characterize nutritional status (C:N:P) and identify nutrient sources ( $\delta^{13}$ C,  $\delta^{15}$ N) supporting HAB phenomena in the IRL (Fig. 3). For macroalgae (Fig. 3D and E), triplicate samples of the two most abundant species were collected by hand, cleaned of epiphytes and debris, rinsed briefly (<5 s) in deionized water to remove excess salt, sorted into three composite replicates per species, and dried at 60 °C for 48 h in a Fisher *Isotemp*<sup>®</sup> laboratory oven. The dried algae were ground to fine powder using a Thompson Scientific Wiley Mini-Mill<sup>®</sup>, and stored in plastic screw top vials. Pre-weighed tissue samples were analyzed for  $\delta^{13}$ C,  $\delta^{15}$ N, total N, and total C at the University of California - Davis's Stable Isotope Facility (SIF) using a PDZ Europa ANCA-GSL elemental analyzer interfaced to a PDZ Europa 20-20 isotope ratio mass spectrometer (IRMS; Sercon Ltd., Cheshire, UK). Samples were combusted at 1000 °C in a reactor packed with chromium oxide and silvered copper oxide. Following combustion, oxides were removed in a reduction reactor (reduced copper at 650 °C). The helium carrier then flowed through a water trap (magnesium perchlorate) and an optional CO2 trap (for N-only analyses). N2 and CO<sub>2</sub> are separated on a Carbosieve GC column (65 °C, 65 mL/min) before entering the IRMS.

Sub-samples of the powdered macroalgae (*n* = 3 replicates/ species/site/sampling event) were analyzed for total P contents at the Nutrient Analytical Services Laboratory, Chesapeake Biological Laboratory, University of Maryland, Solomons, MD. Tissue P was measured following the methodology of Asplia et al. (1976) using a Technicon Autoanalyzer II with an IBM-compatible, Labtronics, Inc. DP500 software data collection system (D'Elia et al., 1997). The %P tissue data, along with %C and %N from the SIF at UC Davis, were used to determine molar C:N:P ratios.

In addition to the macroalgae sampling described above, phytoplankton samples were collected during two unanticipated bloom events during the study, one in the ML and one in CIRL (specifically in the SLE). To characterize nutrient sources supporting a persistent bloom of *Resultor* sp. Ø. Moestrup (Fig. 3A) in the northern end of the IRL, nine phytoplankton samples were collected at three sites in the ML (n = 3 samples/site) in February 2012. At each site, seawater samples were collected in triplicate



**Fig. 3.** Harmful algal bloom species in the Indian River Lagoon: (A) scanning electron micrograph of *Resultor* sp., (B) transmission electron microscopic micrograph of *Aureoumbra lagunensis*, (C) *Lyngbya* sp. (D) *Hydropuntia secunda* (red) and *Chaetomorpha linum* (green), (E) *Caulerpa prolifera*, and (F) *Microcystis aeruginosa* in the St. Lucie Estuary. Photo credits: (A) University of Florida Fisheries and Aquatic Sciences Program, (B) Gobler et al. (2013), (C) Carol Wilson, (D/E) St. Johns River Water Management District, (F) Mark Gavitt, Martin County.

0.25 m below the surface into clean, 1 L HDPE bottles and covered with ice in a dark cooler until return to the HBOI laboratory where they were filtered onto 0.7  $\mu m$  GF/F filters, frozen, and analyzed for  $\delta^{13}C$  and  $\delta^{15}N$  using the methodology described above. Beginning in July 2013, during excessive fresh water releases from Lake

Okeechobee, toxic *Microcystis aeruginosa* (Kützing) Kützing blooms formed in the SLE (Fig. 3F). Three samples of *M. aeruginosa* were collected in August 2013 at Sandsprit Park along Manatee Pocket in the SLE, dried and ground using the methodology described above, and analyzed for  $\delta^{13}$ C,  $\delta^{15}$ N, and C:N:P.

#### 2.3. Species composition in seagrass communities

To characterize each of the 20 IRL sites, an estimate of macroalgae and seagrass abundance (percent cover) was made using five haphazardly tossed 1 m<sup>2</sup> quadrats divided into 100-10 cm<sup>2</sup> cells. Data collection was modified from protocols established by St. Johns River Water Management District (SJRWMD) for their long-term IRL-wide seagrass monitoring project (Morris et al., 2001). Parameters for each haphazard quad included depth, percent drift algae, percent attached algae, and percent seagrass by species. To calculate percent cover, each cell was counted as the parameter of interest either being present or absent. For example, a quad with 36 cells containing seagrass was reported as having 36% cover and drift macroalgae covering 27 cells was reported as 27% cover. Abundant species of macroalgae at each site were photo-documented, collected, and identified to the lowest taxonomic level using local keys (Littler et al., 2008).

### 2.4. Statistical analyses

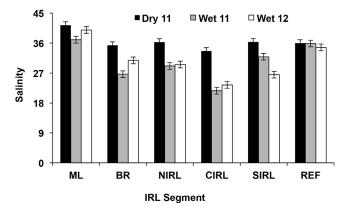
Because of the non-normal distribution of data collected in this study, non-parametric tests were used to determine significance of spatial and temporal differences in measured variables. In IBM SPSS v22, the Kruskal–Wallis test was used to analyze whether or not significant (p < 0.05) differences in measured variables occurred among the IRL segments (ML, BR, NIRL, CIRL, SIRL, and REF) and among the three sampling events (Dry 2011, Wet 2011, and Wet 2012). The Mann–Whitney *U* test was used to identify where the significant differences occurred among the IRL segments and among the three sampling events.

#### 3. Results

Significant temporal and spatial variation in environmental parameters, dissolved nutrients, chlorophyll *a*, stable carbon and nitrogen isotope ratios, molar concentrations of C:N:P, and percent cover of seagrasses occurred during the study.

#### 3.1. Environmental parameters

Salinity showed both significant spatial ( $\chi^2 = 70.553$ ; p < 0.001) and temporal ( $\chi^2 = 55.762$ ; p < 0.001) variation over the two-year study period (Fig. 4). For all three sampling events, salinity along the IRL ranged from 6.0 to 42.9 with an overall IRL mean of  $31.2 \pm 0.6$  and overall REF mean of  $35.5 \pm 0.1$ . The IRL-wide means decreased temporally from  $35.8 \pm 0.8$  during the Dry 2011 sampling to  $29.9 \pm 0.9$  and  $29.9 \pm 0.8$  during the Wet



**Fig. 4.** Mean salinity (±S.E.) in the five IRL segments and the reference sites during the three sampling events in 2011–2012.

2011 and 2012 sampling events, respectively. This temporal variation is also reflected in four of the five IRL segment means where salinity in the dry season was significantly higher than the two wet seasons for BR (p < 0.001), NIRL (p = 0.007), CIRL (p = 0.005), and SIRL (p < 0.001). Salinity also varied spatially in the IRL with the highest mean in ML (39.9 ± 0.5) and lowest in CIRL (25.4 ± 1.6) with other segment means of 31.2 ± 1.0 (BR), 31.8 ± 1.0 (NIRL), 31.5 ± 0.7 (SIRL), and 35.5 ± 0.1 (REF). Water temperature ranged from 18.3 °C in ML to 34.7 °C in the NIRL with an IRL-wide mean of 28.5 ± 0.4 °C and a REF mean of 27.7 ± 0.6 °C during the study. The pH ranged from 6.7 in the CIRL to 8.9 in the NIRL with an IRL-wide mean of 8.0 ± 0.1 and a REF mean of 7.9 ± 0.1. Dissolved oxygen ranged from 54.7% (3.7 mg/L) to 160.0% (9.8 mg/L), where both extremes were in the NIRL. The overall IRL-wide mean was 91.3 ± 5.2% and 6.3 ± 0.4 mg/L and the REF mean was 88.5 ± 4.1% and 5.9 ± 0.5 mg/L.

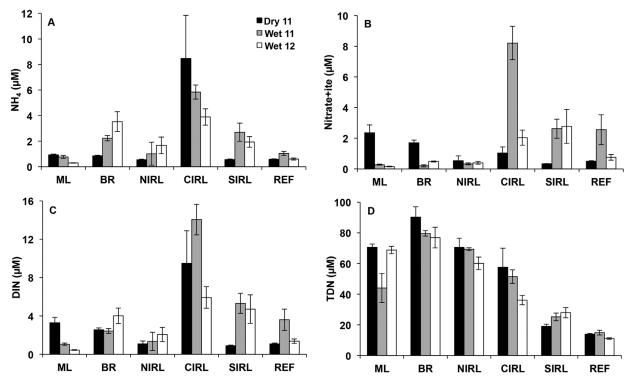
#### 3.2. Dissolved nutrients

Ammonium levels showed significant temporal ( $\chi^2 = 14.834$ ; p = 0.001) and spatial variation ( $\chi^2 = 9.963$ ; p = 0.007) throughout the study period (Fig. 5A). Overall, values ranged from 0.19  $\mu$ M in ML to 34.60  $\mu$ M in the CIRL with an IRL-wide mean of 2.54  $\pm$  0.34  $\mu$ M for all sampling events and an overall REF mean of 0.75  $\pm$  0.07  $\mu$ M. The IRL-wide means varied temporally from 2.22  $\pm$  0.75  $\mu$ M (Dry 2011) to 2.44  $\pm$  0.30  $\mu$ M (Wet 2011) and then down to 2.06  $\mu$ M (Wet 2012). Spatial variation was considerably different with segment means of 0.67  $\pm$  0.07  $\mu$ M (CIRL), 1.73  $\pm$  0.30  $\mu$ M (SIRL), and 0.75  $\pm$  0.07  $\mu$ M (REF). All segments, except ML and NIRL, had significantly higher ammonium values than the reference sites, with *p*-values ranging from p < 0.001 to p = 0.034.

Nitrate/nitrite values only varied spatially ( $\chi^2 = 41.595$ ; p < 0.001) among IRL segments over the study period and were generally lower than the ammonium values (Fig. 5B). During the three sampling events, values ranged from a minimum of  $0.04 \,\mu\text{M}$  to a maximum of  $13.20 \,\mu\text{M}$  with an overall mean of  $1.70\pm0.21~\mu M$  and an overall REF mean of  $1.27\pm0.36~\mu M.$  The IRLwide means had slight temporal variation from 0.95  $\pm$  013  $\mu$ M (Dry 2011) to  $2.70\pm0.45\,\mu M$  (Wet 2011) and then back down to  $1.24\pm0.26~\mu$ M (Wet 2012). Nitrate/nitrite values showed considerable spatial variation with the highest concentrations occurring in the CIRL. Segment means for the three sampling events were  $0.93 \pm 0.26 \ \mu M$  (ML),  $0.80 \pm 0.16 \ \mu M$  (BR),  $0.43 \pm 0.07 \ \mu M$  (NIRL),  $3.76\pm0.63~\mu M$  (CIRL),  $1.91\pm0.45~\mu M$  (SIRL), and  $1.27\pm0.36~\mu M$ (REF). Nitrate/nitrite values were significantly higher in the dry season than in the wet season for ML and BR (p < 0.001), and NIRL (p = 0.044). The reverse occurred in the CIRL, SIRL, and REF, where the Wet 2011 sampling had significantly higher values than the Dry 2011 sampling (*p* < 0.001, *p* = 0.006, and *p* = 0.033, respectively).

Dissolved inorganic nitrogen (DIN) values significantly varied both temporally ( $\chi^2$  = 7.173; p = 0.028) and spatially ( $\chi^2$  = 45.190; p < 0.001) over the study period (Fig. 5C). During the three sampling events, values ranged from 0.26 to 34.90  $\mu$ M with an overall IRL mean of 4.24  $\pm$  0.45  $\mu$ M and an overall REF mean of 2.02  $\pm$  0.42  $\mu$ M. The IRL-wide means varied temporally from 3.17  $\pm$  0.77  $\mu$ M (Dry 2011) to 5.14  $\pm$  0.69  $\mu$ M (Wet 2011) and back down to 3.30  $\pm$  0.46  $\mu$ M (Wet 2012). The CIRL also had the highest DIN concentrations and values showed considerable spatial variation with segment means measuring 1.60  $\pm$  0.30  $\mu$ M (CIRL), 3.64  $\pm$  0.67  $\mu$ M (SIRL), and 2.02  $\pm$  0.42  $\mu$ M (REF). DIN values were significantly different between dry and wet seasons in ML (p < 0.001), the CIRL (p = 0.009), the SIRL (p = 0.004), and the REF sites (p = 0.017).

Total dissolved nitrogen (TDN) values only showed significant spatial variation ( $\chi^2$  = 161.591; p < 0.001; Fig. 5D). Over the three sampling events, TDN values ranged from 7.77 to 152.04  $\mu$ M with



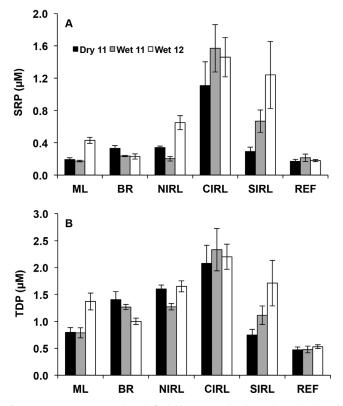
**Fig. 5.** Mean concentrations (±S.E.) of ammonium (A), nitrate-nitrite (B), dissolved inorganic nitrogen (DIN = ammonium + nitrate + nitrite); (C), and total dissolved nitrogen (TDN; D) in the five IRL segments and the reference sites during three sampling events in 2011–2012.

an overall IRL-wide mean of  $53.49 \pm 2.08 \ \mu$ M and an overall REF mean of  $13.32 \pm 0.62 \ \mu$ M. TDN values were similar over the three sampling events with means of  $50.99 \pm 4.26 \ \mu$ M (Dry 2011) to  $46.46 \pm 3.00 \ \mu$ M (Wet 2011) and  $44.13 \pm 2.96$  (Wet 2012). The segment means were  $61.17 \pm 3.99 \ \mu$ M (ML),  $82.30 \pm 3.12 \ \mu$ M (BR),  $66.67 \pm 2.73 \ \mu$ M (NIRL),  $48.45 \pm 4.62 \ \mu$ M (CIRL),  $24.13 \pm 1.51 \ \mu$ M (SIRL), and  $13.21 \pm 0.62$  (REF). All IRL segments, except ML and NIRL, varied significantly from each other ( $p \le 0.001$ ).

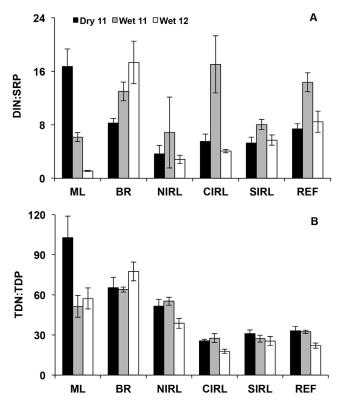
Soluble reactive phosphorus (SRP) concentrations varied significantly both temporally ( $\chi^2 = 7.825$ ; p = 0.020) and spatially ( $\chi^2 = 67.536$ ; p < 0.019) over the study period (Fig. 6A). Overall, values ranged from 0.08 to 4.42  $\mu$ M with an IRL mean of 0.68  $\pm$  0.06  $\mu$ M and a REF mean of 0.19  $\pm$  0.02  $\mu$ M. The IRL-wide segment means varied spatially and increased from 0.44  $\pm$  0.07  $\mu$ M to 0.57  $\pm$  0.09  $\mu$ M to 0.78  $\pm$  0.11 during the in the Dry 2011, Wet 2011, and Wet 2012 sampling events, respectively. IRL segments means for the three sampling events were 0.26  $\pm$  0.03  $\mu$ M (ML), 0.26  $\pm$  0.02  $\mu$ M (BR), 0.40  $\pm$  0.04  $\mu$ M (NIRL), 1.38  $\pm$  0.16  $\mu$ M (CIRL), 0.73  $\pm$  0.16 (SIRL), and 0.19  $\pm$  0.02 (REF). Although there was no significant difference between sampling events in the CIRL, the SRP concentrations were significantly higher in the CIRL when compared with all other segment means, including the reference sites (p < 0.001).

Total dissolved phosphorus (TDP) values only showed significant spatial variation ( $\chi^2 = 108.408$ ; p < 0.001) during the study (Fig. 6B). Over the three sampling events, the TDP values ranged from 0.22 to 5.13  $\mu$ M with an overall IRL-wide mean of 1.48  $\pm$  0.07  $\mu$ M and an overall REF mean of 0.49  $\pm$  0.03  $\mu$ M. IRL-wide means showed little temporal variation and measured 1.23  $\pm$  0.10  $\mu$ M (Dry 2011) to 1.27  $\pm$  0.11  $\mu$ M (Wet 2011) and 1.49  $\pm$  0.12  $\mu$ M (Wet 2012). Spatially, TDP values varied greatly with segment means of 0.98  $\pm$  0.08  $\mu$ M (ML), 1.22  $\pm$  0.05  $\mu$ M (BR), 1.51  $\pm$  0.07  $\mu$ M (NIRL), 2.20  $\pm$  0.19  $\mu$ M (CIRL), 1.19  $\pm$  0.16  $\mu$ M (SIRL), and 0.49  $\pm$  0.03  $\mu$ M (REF). All IRL segments exhibited significantly higher TDP values than the reference sites (p < 0.001) and the CIRL had significantly higher values when compared to all other IRL segments ( $p \le 0.007$ ).

The DIN:SRP ratios showed both significant temporal ( $\chi^2 = 20.698$ ; p < 0.001) and spatial variation ( $\chi^2 = 32.715$ ; p < 0.001) over the study period (Fig. 7A). During the study, DIN:SRP ranged from 0.45 to 51.83 with an overall IRL-wide mean of



**Fig. 6.** Mean concentrations ( $\pm$ S.E.) of soluble reactive phosphorus (SRP; A) and total dissolved phosphorus (TDP; B) in the five IRL segments and the reference sites during the three sampling events in 2011–2012.



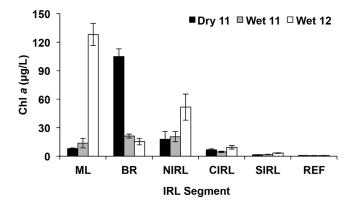
**Fig. 7.** Mean ( $\pm$ S.E.) DIN:SRP (A) and TDN:TDP (B) ratios in the five IRL segments and the reference sites during the three sampling events in 2011–2012.

7.61  $\pm$  0.61 and an overall REF mean of 10.06  $\pm$  0.90. The IRL-wide means showed temporal variation with values from 7.05  $\pm$  0.65 (Dry 2011) to 10.97  $\pm$  1.20 (Wet 2011) and back down to 6.07  $\pm$  0.73 (Wet 2012). DIN:SRP values showed considerable spatial variation with segment means of 7.97  $\pm$  1.55 (ML), 12.84  $\pm$  2.15 (BR), 4.43  $\pm$  0.61 (NIRL), 8.86  $\pm$  1.69 (CIRL), 6.33  $\pm$  0.49 (SIRL), and 10.06  $\pm$  0.90 (REF). The Wet 2011 DIN concentration was significantly higher than that recorded the Dry 2011 for the NIRL (*p* = 0.023), CIRL (*p* = 0.001), the SIRL (*p* = 0.019), and at the REF sites (*p* = 0.001).

TDN:TDP ratios in the IRL also showed significant temporal ( $\chi^2 = 8.612$ ; p = 0.013) and spatial variation ( $\chi^2 = 117.686$ ; p < 0.001) over the study period (Fig. 7B). During the study, TDN:TDP ranged from 10.81 to 176.86 with an IRL-wide mean of 43.71 ± 2.04 and an overall REF mean of 29.20 ± 1.55. The IRL-wide means decreased temporally with values ranging from 47.15 ± 3.82 (Dry 2011) to 41.10 ± 2.13 (Wet 2011), and 36.12 ± 2.79 (Wet 2012). TDN:TDP ratios also showed considerable spatial variation with the higher values occurring in the northern IRL segments. Segment means were recorded as 70.53 ± 7.75 (ML), 69.00 ± 3.17 (BR), 48.55 ± 3.07 (NIRL), 23.70 ± 1.41 (CIRL), 27.94 ± 1.71 (SIRL), and 29.20 (REF). TDN:TDP values were significantly different in ML (p = 0.016), NIRL (p = 0.017), CIRL (p = 0.002), and the REF sites (p = 0.007) during the study.

#### 3.3. Chlorophyll a

Chlorophyll *a* concentrations showed significant spatial variation ( $\chi^2 = 161.924$ ; *p* < 0.001) during the study (Fig. 8). During the three sampling events, chlorophyll *a* values ranged from 0.62 µg/L in the SIRL to 175.33 µg/L in ML with an IRL-wide mean of 21.94 ± 2.66 µg/L and an overall REF mean of 0.75 ± 0.06 µg/L. IRL-wide means varied temporally from 15.55 ± 3.36 µg/L (Dry 2011) to 9.70 ± 1.36 µg/L (Wet 2011) and then 30.23 ± 5.60 µg/L (Wet 2012). Spatially, chlorophyll *a* concentrations varied considerably with



**Fig. 8.** Mean chlorophyll *a* concentrations ( $\pm$ S.E.) in the five IRL segments and the reference sites during the three sampling events in 2011–2012.

segment means of  $49.94 \pm 11.58 \ \mu g/L$  (ML),  $39.52 \pm 8.06 \ \mu g/L$  (BR),  $30.06 \pm 5.16 \ \mu g/L$  (NIRL),  $6.91 \pm 0.77 \ \mu g/L$  (CIRL),  $2.28 \pm 0.19 \ \mu g/L$  (SIRL) and  $0.75 \pm 0.06 \ \mu g/L$  (REF). During the Wet 2012 sampling, chlorophyll *a* concentrations were significantly higher than both the Dry 2011 sampling and the Wet 2011 sampling (p < 0.001). Throughout the study, all IRL segments had significantly higher chlorophyll *a* levels than that documented at the reference sites (p < 0.001).

#### 3.4. $\delta$ 13C, $\delta$ 15N, and C:N:P values of macroalgae and phytoplankton

A total of 44 species of red, green and brown macroalgae were collected in the IRL and analyzed for  $\delta^{13}$ C,  $\delta^{15}$ N and C:N:P contents. The taxa collected included 19 rhodophytes, 17 chlorophytes, and 8 phaeophytes (Table 2).

The  $\delta^{13}$ C values of IRL macroalgae showed significant temporal ( $X^2 = 54.87$ , p < 0.001) and spatial variation ( $X^2 = 17.96$ , p < 0.003) over the two-year study (Fig. 9A). Overall, the  $\delta^{13}$ C values ranged from -33.69% in the SIRL to -9.27% in the BR, with an IRL mean of  $-18.41 \pm 0.22\%$  and a REF mean of  $-19.53 \pm 0.59\%$ . The IRL-wide  $\delta^{13}$ C means varied temporally from  $-17.24 \pm 0.30\%$  (Dry 2011) to  $-20.22 \pm 0.37\%$  (Wet 2011) to  $-18.60 \pm 0.45\%$  (Wet 2012). There was also spatial variation, with overall IRL segment  $\delta^{13}$ C mean values of  $-19.49 \pm 0.35\%$  (ML),  $-16.08 \pm 0.63\%$  (BR),  $-18.38 \pm 0.38\%$  (NIRL),  $-18.27 \pm 0.29\%$  (CIRL) and  $-18.63 \pm 0.72\%$  (SIRL). The ML values were significantly lighter than BR, CIRL, and SIRL values and BR values were significantly heavier than the NIRL, CIRL, and the REF values (Fig. 9A).

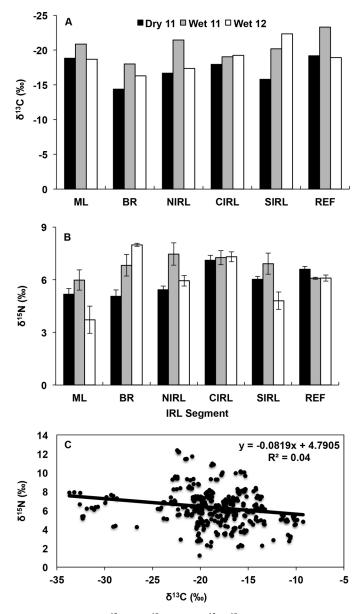
The  $\delta^{15}$ N values of IRL macroalgae showed significant temporal  $(X^2 = 8.71, p = 0.013)$  and spatial variation  $(X^2 = 28.91, p < 0.001)$ over the two-year study (Fig. 9B). Overall, the  $\delta^{15}$ N values ranged from +1.24‰ in ML to +12.37‰ in the NIRL, with an IRL mean of +6.30  $\pm$  0.12‰ and overall REF mean of +6.30  $\pm$  0.09‰. The IRL-wide  $\delta^{15}$ N means varied temporally from +6.06 ± 0.11‰ (Dry 2011) to  $+6.81 \pm 0.22\%$  (Wet 2011) and  $+6.11 \pm 0.18\%$  (Wet 2012). Spatial variation was considerable, with overall IRL segment mean values for  $\delta^{15}$ N of +5.20 ± 0.30‰ (ML), +6.5 ± 0.30‰ (BR), +6.25 ± 0.25‰ (NIRL), +7.11  $\pm$  0.19‰ (CIRL) and +6.00  $\pm$  0.24‰ (SIRL). ML values were significantly lower than all other segments and reference sites. Conversely, CIRL was higher than the NIRL, SIRL and REF sites (Fig. 9B). Based on regression analyses of  $\delta^{15}N$  versus two forms of DIN (ammonium [NH<sub>4</sub>] and nitrate [NO<sub>3</sub>]), there was a trend of macroalgal  $\delta^{15}$ N enrichment with increasing concentrations of both NH<sub>4</sub> and NO<sub>3</sub>. Despite the potential use of both forms of DIN, macroalgae examined during this study were relatively more enriched by NH<sub>4</sub> compared to  $NO_3$  (Fig. 10).

The macroalgae C:N data showed significant temporal and spatial variation in the N-limited status of these benthic algal

NI, ML, ML, ML, ML, ML, ML, ML, ML, ML, ML			Mo La	squito goon		Banan	a River		Noi	thern l	RL			Centr	al IRL				Souther	n IRL		Off	shore ref	rences	
2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2																					t SIRL5 (20)				Coral Cove (24)
	<b>Rhodophyta (red algae)</b> Acanthophora muscoides	ε							×																
	Acanthophora spicifera Botrvocladia occidentalis	1,2,3 1 2												×	×				×			>			
	Bryothamnion seaforthii	2,3						×														< ×	×	×	
<pre>     ***     **</pre>	ryothamnion triquetrum	<del>,</del> ,											:										×		
23 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3	eramium sp. hondria baileyana					×							×												
11222233 2 2 3 3 3 3 3 3 3 3 3 3 3 3 3 3	hondria sp.	2				×																			
	igenea simplex				Y	;		;		;	:	;	:	:		:									
<pre> x x x x x x x x x x x x x x x x x x x</pre>	raciiaria tikvaniae alymenia elongata			×		×		×		×	×	×	×	×		×							×		
	eterosiphonia gibbesii udromutia socurda	1 1 2 2		>				;							;	;						×			
1-22233-33 - 3 3 3 3 3 3 3 3 3 3 3 3 3 3	yunopunna secuna ypnea musciformis	1,2,3		~	×			×		×					×	×		×				×			
x x x x x x x x x x x x x x x x x x x	ypnea spinella	1,3	~						×					×							×				
3 2 2 3 3 3 3 2 5 3 3 3 3 3 3 4 × × × × × × × × × × × × ×	turencia filiformis turencia poiteaui	- E		~	×		×	×																	×
<pre> 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2</pre>	olieria filiformis wridia filamentosa	. 1	>													>						×			
2 2 3 3 3 3 3 3 3 3 3 3 3 3 3		-	<													×									
1       3	nloropnyta (green algae) Prahularia of farlowii	c						*																	
23 23 23 23 23 24 25 2 2 2 2 2 2 2 2 2 2 2 2 2	cetabularia schenckii	2 2			×			<																	
Jeffal       123         Jeffal       123         123       2333         123       2333         123       2333         123       2333         123       2333         123       2333         123       233         123       233         133       233         133       233         133       233         1430       233         133       233         133       233         1430       233         153       233         1430       233         153       233         1430       233         153       233         153       233         1430       23         153       23         1430       24         153       25         153       25         153       25         153       25         153       25         153       25         153       25         153       25         154       25         155	etabularia sp.	2,3			×																				
	uulerpa mexicana nulerna nrolifera	1 1 2 3							>							×	×	>	>	>		>			
pettac 1 3 3 3 4 5 3 3 5 3 3 5 3 5 3 5 5 5 5 5 5	ulerpa racemosa	1,2							¢									¢	<	¢		< ×		×	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	ulerpa racemosa v. peltata	1													:		;		:	:	:				×
z z z z z z z z z z z z z z z z z z z	auterpa sertutariotaes aulerpa verticillata	2,3													×		×		××	××	×				
z z z z z z z z z z z z z z z z z z z	haetomorpha gracilis											×													
<pre>     fige         x x</pre>	adophora laetevirens Iadonhora nrolifera	2,3						×			×														
2 2 2 3 3 3 3 3 3 3 3 3 3 3 3 3	alimeda incrassata	5																		×					
x x x x x x x x x x x x x x x x x x x	lva flexuosa Iva intestinalis	2 1 2 3	×			>			>	>	>		>	>											×
X X X X X X X X X X X X X X X X X X X	lva rigida	2				<			<	<	<		<	<											×
alge)	lva sp.		2		×	×														×					
x x x x x x x x x x x x x x x x x x x	haeophyta (brown algae) ictuortaris instii	6																					>		
× × ×	Dictyota cervicornis	n —																			×		<	×	
z z z z x x x x x x x x x x x x x x x x	ictyota sp.	ε																						×	
< < <	lincksia mitchelliae incksia sandriana	7 7	×														>	>							
1 ×	adina gymnospora	1															<	<							×
	Padina sp.	1																					*		

Table 2
 Macroalgae collected during the Dry 2011, Wet 2011, and Wet 2012 sampling seasons at 20 sites in the Indian River Lagoon and 4 adjacent nearshore reef Reference sites. Sampling Event Codes: (1) May 2011, (2) October 2011, (3)
 October 2012.

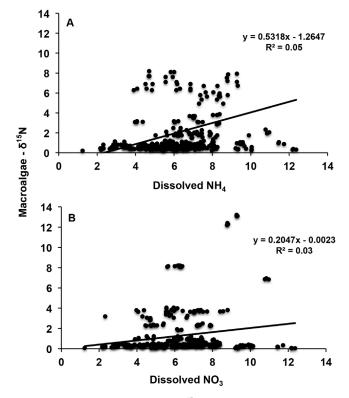
91



**Fig. 9.** Mean (±S.E.)  $\delta^{15}N$  (A),  $\delta^{13}C$  (B) and  $\delta^{15}N/\delta^{13}C$  regression bi-plot (C) for macroalgae collected from each of the five IRL segments and the reference sites during the three sampling events in 2011–2012.

blooms during the study (Table 3). Over the three sampling events, the C:N values ranged from 9.28  $\pm$  0.23 in the CIRL to 42.40  $\pm$  1.50 in BR, with an overall IRL mean of 15.87  $\pm$  0.44 and overall REF mean of 17.02  $\pm$  0.73. The IRL-wide C:N means decreased temporally through the study from 17.74  $\pm$  0.62 (Dry 2011) to 15.55  $\pm$  0.78 (Wet 2011) and 14.36  $\pm$  0.53 (Wet 2012). Spatial variation in C:N among IRL segments was considerable, with overall segment mean values ranging 10.54  $\pm$  0.35 in the CIRL during the Wet 2011 sampling to 23.94  $\pm$  3.76 in the BR during the Dry 2011 sampling.

The macroalgae C:P data showed significant temporal and spatial variation in the degree of P-limitation in these benthic algal blooms during the study, with generally higher values in the three northern IRL segments (Table 3). Over the three sampling events, the C:P values ranged from  $102.42 \pm 2.82$  in the SIRL to  $3281.6 \pm 813.78$  in ML, with an overall IRL mean of  $698.9 \pm 43.6$  and overall REF mean of  $505.96 \pm 199.1$ . The IRL-wide C:P means varied temporally through the study from  $662.7 \pm 45.4$  (Dry 2011) to  $590.6 \pm 55.0$  (Wet 2011) and  $737.6 \pm 87.7$  (Wet 2012). Spatial



**Fig. 10.** Regression analysis of macroalgal  $\delta^{15}$ N versus dissolved concentrations of ammonium (A) and nitrate (B) for the five IRL segments sampled in 2011–2012.

variation in C:P among IRL segments was also considerable, with overall segment mean values ranging from  $281.8 \pm 18.4$  in the CIRL during the Dry 2011 sampling to  $3281.6 \pm 813.78$  in the ML during the Wet 2012 sampling.

Similar to the C:P ratio, the N:P data showed significant temporal and spatial variation in the degree of P-limitation in benthic algal blooms during the study, with higher values in the three northern segments of the IRL (Table 3). Over the three sampling events, the N:P values ranged from  $11.24 \pm 0.27$  in the SIRL to  $151.50 \pm 36.12$  in ML, with an overall IRL mean of  $40.62 \pm 1.85$  and an overall REF mean of  $31.31 \pm 1.21$ . The IRL-wide N:P means increased temporally through the study from  $35.27 \pm 1.65$  (Dry 2011) to  $35.61 \pm 2.40$  (Wet 2011) and  $47.59 \pm 3.93$  (Wet 2012). Spatial variation in N:P among IRL segments was considerable, with overall segment mean values ranging from  $20.53 \pm 1.01$  in the CIRL during the Dry 2011 sampling to  $151.50 \pm 36.12$  in the ML during the Wet 2012 sampling.

The  $\delta^{13}$ C and  $\delta^{15}$ N values for phytoplankton collected throughout ML during the final stages of the "superbloom" in February 2012 averaged  $-22.22 \pm 0.07$  and  $+3.49 \pm 0.08$ , respectively. The *Microcystis aeruginosa* collected from Manatee Pocket in the CIRL's SLE in August 2013 had  $\delta^{13}$ C and  $\delta^{15}$ N values of  $-24.30 \pm 0.04$  and  $+8.59 \pm 0.06$ , respectively. The C:N, C:P and N:P values for *M. aeruginosa* were  $7.39 \pm 0.10$ ,  $240.85 \pm 6.49$ , and  $32.65 \pm 0.62$ , respectively.

#### 3.5. Species composition in seagrass communities

The percent cover of seagrasses at the 20 IRL study sites ranged from 0 to 100 percent with an overall mean of  $46.67 \pm 2.63$ . There was a significant decline in seagrass cover among the three sampling events ( $X^2 = 9.001$ , p = 0.011). Significantly more seagrass cover was documented in the Dry 2011 ( $57.11 \pm 4.54$ ) season when compared to the Wet 2011 ( $43.08 \pm 4.55$ ) and Wet 2012 ( $39.07 \pm 4.41$ ) seasons (p = 0.018 and p = 0.006, respectively). Significant differences were

	reef	
	earshore	
	cent ne	
	e adja	
	long the adj	
	ites alc	
	ence si	
	Refer	
	on and 4	
	Lagoon	
	River L	
	Indian I	
	the Ir	
	within	
	) sites	
	s at 20	
	and Wet 2012 seasons	
	2012	
	d Wet	
	÷	
	. Wet 201	
	/ 2011	
	in the Dry	
	ed in th	
	collect	
	algae	
	macro	
	) from	
	(±S.E.	
	ratios	
,	C:N:P	
	Mean	

Segment	Site #	Site		Dry 2011			Wet 2011			Wet 2012	
			C: N	C:P	N:P	C:N	C:P	N:P	C:N	C:P	N:P
Mosquito	1	ML1	$11.70\pm1.29$	$268.96 \pm 15.72$	$23.79 \pm 1.50$	$22.40 \pm 1.42$	$341.63 \pm 77.25$	$14.50 \pm 2.56$	I	I	I
Lagoon	2	ML2	$15.81\pm0.45$	$1042.93 \pm 34.46$	$66.23 \pm 2.21$	$18.03\pm1.31$	$595.53 \pm 98.60$	$32.66 \pm 2.96$	I	I	I
	e i	ML3	$17.59 \pm 1.44$	$1413.52 \pm 197.17$	$78.97 \pm 5.64$	$17.23 \pm 3.63$	$1497.25 \pm 628.67$	$80.31 \pm 17.12$	$21.34 \pm 0.86$	$3281.56 \pm 813.78$	$151.50 \pm 36.12$
	Segment Mean±SE	Mean ± SE	$15.03 \pm 0.86$	$908.47 \pm 131.63$	$56.33 \pm 6.05$	$20.02 \pm 1.29$	$694.01 \pm 200.73$	$35.49 \pm 9.00$	$21.34 \pm 0.86$	$3281.56 \pm 813.78$	$151.50 \pm 36.12$
Banana	4	BR1	$\textbf{42.40} \pm \textbf{1.50}$	$2194.70 \pm 45.79$	$51.95 \pm 1.31$	$12.68\pm0.46$	$211.55 \pm 13.89$	$16.69\pm0.55$	$21.05 \pm 0.55$	$1005.89 \pm 38.50$	$47.99 \pm 2.66$
River	J.	BR2	$\textbf{27.29} \pm \textbf{5.51}$	$880.35 \pm 106.07$	$36.56 \pm 5.31$	$\textbf{35.04} \pm \textbf{8.03}$	$1129.14 \pm 165.35$	$36.58 \pm 3.95$	$9.17\pm0.38$	$418.77 \pm 21.02$	$46.31 \pm 3.48$
	9	BR3	$11.35\pm0.56$	$344.25 \pm 41.14$	$29.98 \pm 2.32$	$11.50\pm0.12$	$610.36 \pm 36.39$	$53.21 \pm 3.52$	$14.38 \pm 0.14$	$941.94 \pm 23.93$	$65.65 \pm 1.52$
	Segment Mean $\pm$ SE	Mean ± SE	$23.94 \pm 3.76$	$928.78 \pm 186.10$	$37.01 \pm 3.07$	$23.57 \pm 5.16$	$770.05 \pm 140.76$	$35.76 \pm 4.40$	$13.44 \pm 1.49$	$696.34 \pm 85.11$	$51.57 \pm 3.04$
Northern	7	NIRL1	$\textbf{29.82} \pm \textbf{2.62}$	$\bf 2036.25 \pm 111.78$	$\textbf{74.12} \pm \textbf{7.25}$	$20.79 \pm 0.86$	$\bf 2039.54 \pm 188.58$	$99.86 \pm 9.68$	$17.10\pm1.04$	$1411.41 \pm 173.69$	$\textbf{80.98} \pm \textbf{7.00}$
IRL	8	NIRL2	$16.32 \pm 0.11$	$779.85 \pm 19.83$	$\textbf{47.87}\pm\textbf{0.90}$	$19.74 \pm 0.06$	$925.73 \pm 224.74$	$46.99 \pm 1.40$	$10.71\pm0.62$	$648.25 \pm 115.25$	$57.76 \pm 6.88$
	6	NIRL3	$17.48 \pm 0.34$	$230.42 \pm 13.16$	$13.21\pm0.71$	$10.29 \pm 0.30$	$262.89 \pm 9.40$	$25.73 \pm 1.03$	$14.11 \pm 1.07$	$305.09 \pm 23.63$	$\textbf{21.68} \pm \textbf{0.40}$
	10	NIRL4	$11.42\pm0.67$	$265.56 \pm 13.16$	$\textbf{23.21}\pm\textbf{0.68}$	$10.86 \pm 0.64$	$300.62 \pm 43.05$	$27.06 \pm 2.40$	$9.57\pm0.55$	$233.28 \pm 12.92$	$\textbf{24.48}\pm\textbf{0.43}$
	Segment Mean $\pm$ SE	Mean $\pm$ SE	$21.49 \pm 1.90$	$1101.87 \pm 173.82$	$46.35\pm6.09$	$14.97 \pm 1.03$	$937.14 \pm 169.15$	$53.10 \pm 7.32$	$13.75 \pm 0.72$	$839.88 \pm 114.32$	$55.87 \pm 5.35$
Central	11	CIRL1	$20.65 \pm 0.71$	$284.32 \pm 26.19$	$13.75\pm1.02$	I	I	I	$11.21 \pm 0.37$	$199.23 \pm 9.60$	$17.79 \pm 0.26$
IRL	12	CIRL2	$9.28\pm0.23$	$206.07 \pm 19.54$	$22.28 \pm 2.06$	$11.24 \pm 0.14$	$187.93 \pm 12.63$	$16.79 \pm 1.29$	I	I	I
	13	CIRL3	$11.31 \pm 0.52$	$203.86 \pm 18.99$	$17.82 \pm 1.02$	$11.04 \pm 0.04$	$229.99 \pm 7.70$	$20.88 \pm 0.64$	$10.03 \pm 0.37$	$213.98 \pm 15.35$	$21.30 \pm 1.10$
	14 1	CIKL4	$\frac{1100}{100} \pm \frac{1100}{100} \pm 10$	$335.81 \pm 38.70$	$22.30 \pm 1.65$	$10.49 \pm 0.96$	$298.06 \pm 65.7$	$26.71 \pm 3.82$	$14.21 \pm 0.83$	3/2.60±4/.31	$20.80 \pm 1.31$
	c1		14.92 ± 0.48	418.15 ± 55.29	$50.1 \pm 73.00$	9.99 ± 0.41	239.29 ± 40.74	24.07 ± 5.91			
	Segment Mean ± SE	Mean ± SE	$14.00 \pm 0.76$	<b>281.84</b> ± 18.42	10.1 ± £ c.02	$10.54 \pm 0.35$	248.77 ± 26.07	<b>23.22 ± 1.92</b>	$11.94 \pm 0.62$	$2/4.48 \pm 28.58$	22.39 ± 1.17
Southern	16	SIRL1	$14.58 \pm 0.28$	$367.65 \pm 48.92$	$25.34 \pm 3.37$	$\textbf{9.13}\pm\textbf{0.03}$	$102.42 \pm 2.82$	$11.24\pm0.27$	I	I	I
IRL	17	SIRL2	$20.15 \pm 1.96$	$352.03 \pm 63.07$	$19.67 \pm 4.91$	$9.72 \pm 0.07$	$161.34 \pm 17.27$	$16.66\pm1.91$	I	I	I
	18	SIRL3	$15.10 \pm 0.22$	$597.66 \pm 38.90$	$39.60 \pm 2.24$	$11.78\pm0.35$	$370.71 \pm 12.19$	$31.57 \pm 0.64$	$10.11\pm0.37$	$263.37 \pm 31.65$	$26.50 \pm 3.68$
	19	SIRL4	$17.19\pm0.83$	$575.48 \pm 70.49$	$34.67\pm5.39$	$20.48 \pm 3.59$	$932.70 \pm 44.62$	$49.42 \pm 7.22$	$10.97\pm0.44$	$426.67 \pm 22.67$	$39.72 \pm 3.00$
	20	SIRL5	$20.28 \pm 1.54$	$630.28 \pm 50.69$	$31.13 \pm 0.72$	$14.81\pm0.35$	$372.46 \pm 20.79$	$25.16 \pm 0.83$	I	I	I
	Segment Mean $\pm$ SE	Mean ± SE	$17.46 \pm 0.67$	$504.62 \pm 32.02$	$30.08 \pm 2.02$	$13.34 \pm 1.18$	$413.88 \pm 68.57$	$28.75 \pm 3.34$	$10.63 \pm 0.31$	$361.35 \pm 27.86$	$34.43 \pm 2.83$
Reference	21	Ambersand	$15.63\pm1.38$	$509.80 \pm 55.40$	$33.61 \pm 2.97$	$10.27 \pm 0.24$	$324.27 \pm 52.99$	$31.75 \pm 5.37$	$18.49 \pm 3.19$	$396.60 \pm 27.04$	$25.40 \pm 2.98$
	22	Pepper Park	$15.00\pm1.56$	$476.30 \pm 65.44$	$31.24 \pm 1.13$	$13.86 \pm 1.13$	$315.42 \pm 93.07$	$24.12 \pm 8.04$	$23.10 \pm 0.66$	$435.88 \pm 27.59$	$18.86\pm0.86$
	23	Bathtub	$16.85 \pm 1.54$	$547.96 \pm 49.99$	$32.76 \pm 1.69$	$13.37 \pm 0.32$	$480.98 \pm 38.80$	$36.12 \pm 3.01$	$15.51\pm0.16$	$598.37 \pm 61.59$	$38.71 \pm 4.13$
	24	Coral Cove	$24.32 \pm 2.02$	$852.17 \pm 86.94$	$35.63 \pm 2.92$	$26.15 \pm 0.29$	$697.27 \pm 29.16$	$26.72 \pm 2.07$	$11.31\pm0.37$	$548.56 \pm 60.55$	$49.15\pm3.85$
	Segment <b>P</b>	Segment Mean $\pm$ SE	$17.32 \pm 0.99$	$572.90 \pm 39.67$	$33.39 \pm 1.47$	$15.46 \pm 0.14$	$455.59 \pm 32.45$	$30.63 \pm 3.29$	$18.33 \pm 1.59$	$458.36 \pm 23.94$	$28.82 \pm 2.71$
	Seasonal Mean±SE	Mean + SE	$17.74 \pm 0.62$	662 69 + 45 41	35 77 + 1 65	15 55 ± 0 78	590.60 + 55.01	35.61 + 2.40	11 36 + 0 53	727 56 + 87 74	47 59 + 3 93

also documented in the seagrass cover within the five segments ( $X^2$  = 5.510, p < 0.001) where the mean coverage over the extent of the study was 56.09 ± 6.93 (ML), 32.70 ± 6.78 (BR), 39.15 ± 6.17 (NIRL), 34.29 ± 4.80 (CIRL), and 67.76 ± 4.45 (SIRL).

The coverage of four dominant species of seagrass (Halodule wrightii Ascherson, Halophila johnsonii N.J. Eiseman, Syringodium filiforme Kützing, and Thalassia testudinum K.D. Koenig) varied both temporally and spatially over the study period (Fig. 11). Overall, H. wrightii, located in all five segments of the IRL, was the most abundant seagrass species in the system with percent cover of  $38.78 \pm 2.57$ . The next most abundant seagrass species was S. filiforme, which was located at study sites in all IRL segments except ML and BR, and had a percent cover of  $6.02 \pm 1.30$ . The federally protected seagrass, H. johnsonii, and T. testudinum were only present at study sites in the CIRL and SIRL where they had a percent covers of  $5.35 \pm 1.11$  and  $3.00 \pm 0.91$ , respectively. Temporally, the mean percent cover of H. wrightii in Dry 2011, Wet 2011, and Wet 2012 was  $48.00 \pm 4.50$ ,  $39.59 \pm 4.52$ , and  $28.32 \pm 4.10$ , respectively. *S. filiforme* percent cover decreased from 6.12  $\pm$  2.13 (Dry 2011) to 1.82  $\pm$  1.28 (Wet 2011) then increased again to 9.88  $\pm$  2.89 (Wet 2012). Similar to H. wrightii, H. johnsonii continuously decreased from 8.53  $\pm$  2.38 (Dry 2011) to 5.32  $\pm$  2.16 (Wet 2011) to 2.04  $\pm$  0.67 (Wet 2012). T. *testudinum* percent cover was lowest in Dry 2011 ( $1.90 \pm 1.15$ ) then increased in Wet 2011 (5.04  $\pm$  2.10) and then decreased again in Wet 2012 (2.23  $\pm$  1.39). Spatially, the mean percent cover of *H. wrightii* over the study period ranged from  $29.00 \pm 4.53$  in the CIRL to  $56.09 \pm 6.93$  in ML. In the segments where S. *filiforme* was present (NIRL, CIRL, and SIRL), percent cover ranged from  $3.63 \pm 1.90$  in the CIRL to  $13.43 \pm 3.68$  in the SIRL. The mean cover of *H. johnsonii* in the CIRL and SIRL was  $2.71 \pm 1.26$  and  $18.90 \pm 3.89$ , respectively. Similarly, the mean percent cover of T. testudinum in the CIRL and SIRL was 5.57  $\pm$  2.33 and 6.26  $\pm$  2.64, respectively.

Attached and drift macroalgae were also documented within the seagrass communities during all three samplings. The overall mean percent cover of attached and drift algae, respectively, was 27.58  $\pm$  3.52 and 3.46  $\pm$  1.79 (Dry 2011), 5.77  $\pm$  1.60 and 0.88  $\pm$  0.48 (Wet 2011), and 9.59  $\pm$  2.40 and 6.05  $\pm$  1.71 (Wet 2012). Spatial variation in percent cover of attached and drift algae, respectively, was observed amongst the regions with 5.40  $\pm$  1.86 and 0.00  $\pm$  0.00 (ML), 9.06  $\pm$  2.99 and 0.00  $\pm$  0.00 (BR), 17.80  $\pm$  5.95 and 0.00  $\pm$  0.00 (NIRL), 18.83  $\pm$  5.81 and 2.68  $\pm$  1.89 (CIRL), 37.08  $\pm$  6.85 and 0.92  $\pm$  0.28 (SIRL) during the Dry 2011 season. In the Wet 2011 season, the variation in percent cover of attached and drift macroalgae, respectively, was 1.27  $\pm$  1.07 and 4.00  $\pm$  2.89 (ML), 0.00  $\pm$  0.00 and 0.00  $\pm$  0.00 (BR),

 $6.05 \pm 3.44$  and  $0.20 \pm 0.20$  (NIRL),  $4.04 \pm 1.53$  and  $0.04 \pm 0.04$  (CIRL), and  $12.28 \pm 4.99$  and  $0.76 \pm 0.46$  (SIRL). In the Wet 2012 season, the percent cover of attached and drift algae varied respectively,  $0.00 \pm 0.00$  and  $4.80 \pm 2.48$  (ML),  $8.87 \pm 6.02$  and  $1.87 \pm 1.00$  (BR),  $14.16 \pm 5.53$  and  $10.04 \pm 3.80$  (NIRL),  $5.52 \pm 2.24$  and  $0.00 \pm 0.00$  (CIRL),  $15.75 \pm 7.79$  and  $12.1 \pm 6.37$  (SIRL).

#### 4. Discussion

# 4.1. Nitrogen enrichment drives phosphorus-limited phytoplankton blooms

This study represents the first IRL-wide water quality assessment that integrates water column nutrient monitoring with macroalgae "bio-observatories" and seagrass surveys to identify nutrient sources and gauge the type and degree of N versus P limitation. Macroalgae in the IRL are known to respond to increased N availability with increased growth (Lapointe and Ryther, 1979) and excessive biomass has been symptomatic of accelerating eutrophication for several decades (Bricker et al., 2007). The timing of our study was serendipitous as it coincided with the end of a multi-year drought, reduced abundance of drift macroalgae (personal observations, SJRWMD et al., 2012), and development of unprecedented phytoplankton blooms in the northern segments of the IRL. This began with the development of the "superbloom" ( $\sim$ 130 µg/L) in the BR, NIRL and ML that was comprised predominantly of the microflagellate, Resultor sp., Fig. 3(A) and persisted between March 2011 and February 2012, with the highest concentrations between April and November 2011 (Phlips et al., 2014). Seagrass loss (~45% or ~31,600 acres by June 2011; SJRWMD, unpublished data) immediately following the onset of the superbloom during the first year of our study was substantial and was reflected in our Wet 2011 seagrass survevs.

The superbloom was followed by an unprecedented brown tide in July 2012 comprised of the pelagophyte *Aureoumbra lagunensis* D.A. Stockwell, DeYoe, Hargraves and P.W. Johnson in the southern ML and NIRL (Gobler et al., 2013), which caused the highest chlorophyll *a* values of our study. Coastal waters susceptible to brown tides are relatively warm, shallow, and hypersaline, with little tidal exchange (high residence times; Gobler and Sunda, 2012); however, these environmental conditions have been commonly seen in these segments of the IRL when such severe phytoplankton blooms have not occurred (Briel et al., 1973). In August 2012, we

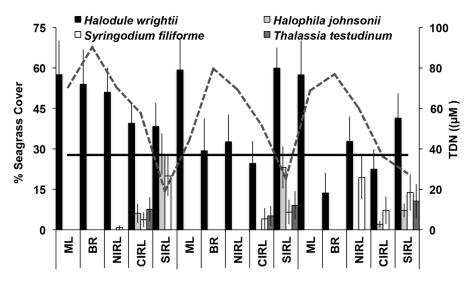


Fig. 11. Mean seagrass cover (±S.E.) and total dissolved nitrogen (TDN) by species in each of the five IRL segments during the three sampling events in 2011–2012. The dashed line denotes TDN concentrations during the study. The solid black line represents the total nitrogen threshold above which *Thalassia testudinum* becomes stressed (Lapointe et al., 1994).

observed >130 µg/L chlorophyll *a* while others reported up to 196 µg/L chlorophyll *a* and  $3 \times 10^6$  cells/L in ML, the epicenter of the bloom (Gobler et al., 2013). A partial die-back occurred in September 2013 following the passing of tropical storm Isaac. In April 2013, a smaller *A. lagunensis* bloom re-emerged in the same area (SJRWMD, 2013). These IRL events were similar to the prolonged brown tides in the 1990s along coastal lagoons in Texas (Buskey et al., 2001; Gobler and Sunda, 2012) that similarly resulted in seagrass die-off due to reductions in light (Onuf, 1996).

Although the long residence time of water in the three northern IRL segments has long been recognized as a physical factor contributing to bloom development (Briel et al., 1973), our nutrient data indicate that these northern segments were also highly enriched in TDN (61-82 µM) at levels well above FDEP TMDL targets ( $\sim$ 50  $\mu$ M) with correspondingly high TDN:TDP ratios (49:1– 71:1) during these phytoplankton bloom events. The more highly flushed CIRL and SIRL had lower TDN concentrations (24-48 µM), TDN:TDP ratios (24–29) and chlorophyll a concentrations and were not impacted by severe phytoplankton blooms or seagrass die-off during our study. These observations are consistent with previous studies (Sigua et al., 2000; Phlips et al., 2010), however, our interpretations of how the stoichiometry impacts the system is new. Although high N:P ratios generally indicate P-limitation, a variety of phytoplankton can have high growth rates at high N:P ratios (>70) while experiencing strong P-limitation (Terry et al., 1985). For example, Aureoumbra lagunensis, which formed persistent and damaging blooms in Laguna Madre, TX, is capable of growing under a wide range of N:P ratios (Liu et al., 2001) and actually formed dense blooms when the water column N:P ratio increased to high levels (~140; Rhudy et al., 1999). Similarly, the cyanobacterium Synechococcus spp., which formed severe and damaging blooms in Florida Bay between 1991 and 1996 following increased freshwater flows and N-loading from Everglades runoff (Brand, 2002; Lapointe and Barile, 2004), can outcompete other phytoplankton species under high N:P ratios and P-limited conditions (Lavrentyev et al., 1998; Richardson, 2009).

These data and observations contrast with the conclusions of Sigua et al. (2000) that algal blooms in the IRL are most likely to develop in the CIRL and SIRL where "a TN/TP ratio  $\leq$ 10 appears to favor algal blooms, especially blue-green algae, which are capable of fixing atmospheric N." Indeed, the cyanobacterium Microcystis aeruginosa formed blooms in the SLE during June 2005 and July 2013 following heavy freshwater releases from Lake Okeechobee and runoff from the surrounding SLE basin (Fujimoto et al., 1997; Phlips et al., 2012; Lapointe et al., 2012). Although a decrease in the water column N:P ratio accompanied the M. aeruginosa blooms (Lapointe et al., 2012; Parrish, 2014), high P (relative to N) loading is not a universal "trigger" for cyanobacterial blooms (Paerl and Otten, 2013). The "P only" paradigm assumed that N<sub>2</sub> fixation could supply all of the required N, a viewpoint that needs revision to account for many cyanobacterial blooms that result from Nenrichment and occur at high N:P ratios (Paerl and Fulton, 2006). Toxic strains of *M. aeruginosa*, which produced the hepatotoxin microcystin, were documented in the SLE during both 2005 and 2013, resulting in numerous public warnings from the Department of Health cautioning residents and visitors to avoid recreational use of the SLE, IRL, and nearshore reefs adjacent to St. Lucie Inlet (Phlips et al., 2012; FDEP, unpublished data). Furthermore, a variety of HABs show increased toxin production under Nenrichment and increased P-limitation (Donald et al., 2011; Dolman et al., 2012; Hardison et al., 2013), which readily explains why toxins could be especially problematic in the northern segments of the IRL, where unusual mass mortality events, involving manatees, dolphins, and pelicans, developed in 2012 and 2013 following these unprecedented bloom events (SJRWMD, 2013; Wines, 2013; NOAA, 2014).

#### 4.2. Freshwater runoff transports dissolved nutrients into the IRL

Multiple factors and nutrient sources likely worked in synergy to generate the unprecedented phytoplankton blooms in the northern segments of the IRL during our study. Prior to the blooms, the adjacent IRL watersheds experienced a multi-year drought that culminated with a "first flush" during spring and early summer 2011 (Fig. 2). These northern segments, which had high TDN:TDP ratios and were P-limited, experienced a spike in P during the spring 2011 following the end of the drought with increasing rainfall (SIRWMD et al., 2012). However, we also observed relatively little biomass of benthic macroalgae in the IRL during the study, possibly the result of reduced nutrient loading associated with the antecedent drought. In addition to the spike in nutrient loading from increased stormwater runoff, ash produced during a controlled burn in July 2011 within Merritt Island National Wildlife Refuge may also have contributed a pulse of P that would likely have fueled the bloom. Regardless of the source, the pulses of P into the northern segments of the IRL, where minimum macroalgal biomass was available to assimilate nutrients, culminated in the unprecedented phytoplankton blooms that resulted in a 60% decline in seagrass in the northern IRL (SIRWMD, 2013). Below we describe the spatial and temporal patterns among salinity, dissolved N and P concentrations, N:P ratios, chlorophyll a, as well as  $\delta^{13}$ C and  $\delta^{15}$ N values for macroalgae and phytoplankton HABs that supports our conclusions.

Overall, salinity generally decreased through the course of our two-year study, indicating that the observed algal blooms were a response to increasing land-based nutrient inputs. The lowest salinity values during our study were in the CIRL (mean 25.37), the segment of the IRL that receives considerable freshwater discharges from three drainage canals in Indian River County, as well as the St. Sebastian River, the C-54 canal, and several creeks in southern Brevard County (Fig. 1). Estimated freshwater flows into this region of the IRL are  $\sim$ 500,000,000 m<sup>3</sup> yr<sup>-1</sup>, which is dominated (168,039,748  $m^3 yr^{-1}$ ) by the St. Sebastian River (Sigua and Tweedale, 2003). Not surprisingly, we found the highest concentrations of ammonium (6.60  $\mu$ M), nitrate/nitrite (3.76  $\mu$ M), DIN (9.83  $\mu$ M), SRP (1.38  $\mu$ M), and TDP (2.20  $\mu$ M) associated with these low salinity values in the CIRL. These findings are in general agreement with previous IRL water quality assessments, which report the highest TN (765,211 kg N/yr) and TP (87,978 kg/yr) nonpoint source loadings occurred in this segment of the IRL (Sigua and Tweedale, 2003).

Despite the large freshwater inflows and high dissolved inorganic N and P concentrations in the CIRL, the phytoplankton blooms occurred in the NIRL, BR and ML where dissolved inorganic nutrients were relatively low but TDN concentrations and TDN/ TDP ratios were high. We attribute this disconnect between the IRL segment with the highest dissolved inorganic nutrient concentrations and that with the most severe phytoplankton bloom development to several factors. First, the CIRL has a much shorter water residence time as a result of higher tidal flushing via the Sebastian and Ft. Pierce inlets that would limit phytoplankton bloom development despite high DIN and SRP concentrations (Lu and Gan, 2014); the longer water residence times in the northern segments would support greater development of microbial biomass and preferential scavenging of inorganic nutrient pools (Lucas et al., 1999). Second, the longer water residence time and microbial biomass in the NIRL, ML and BR allows for a higher degree of autotrophic and heterotrophic nutrient cycling, which would tend to not only deplete the more biologically available DIN and SRP, but also elevate DON and DOP concentrations over time (Bronk et al., 1998; Tyler et al., 2001). Third, urbanization has correlated with increases in macroalgae in the IRL, as is the case in many other estuaries and coastal waters (Lapointe et al., 1994;

Valiela et al., 1992). Historically, macroalgae were a minor component of seagrass communities in the IRL (Benz et al., 1979; Gilbert and Clark, 1981; Virnstein and Carbonara, 1985), but in recent years their levels have increased to three times the seagrass biomass throughout the IRL as a whole (Hall et al., 2001; Riegl et al., 2005). These macroalgal blooms are a response of the shallow estuary to nutrient enrichment and can cause reduced light, hypoxia, fragmentation and die-off of seagrasses (Lapointe et al., 1994; Valiela et al., 1997). Because macroalgae are known to assimilate DIN and release DON (Tyler et al., 2001), these benthic macroalgal blooms would also result in cumulative build-up of DON in the IRL over time.

The mean C:N:P ratios of macroalgae in the IRL indicated that growth of these shallow primary producers is potentially limited by both N and P. The overall mean C:N ratio was  $\sim$ 17, with higher values, up to 42, in the northern segments of the IRL. The lowest C:N ratios occurred in the CIRL where the highest DIN occurred, with the highest values in the NIRL, BR, and ML. Because C:N ratios >12 indicate increasing N-limitation (D'Elia and DeBoer, 1978; Lapointe et al., 1984), the IRL values, especially in the NIRL, BR and ML, indicate significant N-limitation and supports previous experimental enrichment studies (Lapointe and Ryther, 1979; Hanisak, 1990). As N-loading generally increased with increasing rainfall and decreasing salinity through the study, the increased N availability was reflected in decreasing C:N ratios of macroalgae. The mean C:P and N:P ratios were 495 and 31, respectively, indicating significant co-limitation by P. However, overall higher C:P and N:P ratios occurred in the northern IRL segments, indicating stronger P-limitation in this region of the IRL. These high macroalgae C:P and N:P ratios mirror the high seawater TDN/ TDP ratios in this part of the IRL, and could indicate cumulative inputs of groundwater with high N:P ratios. Preferential scavenging of P by soils results in increasing N:P ratios along the flow path of OSTDS plumes in groundwaters and has been linked to high N:P ratios and P-limitation in downstream surface waters (Lapointe et al., 1990; Weiskel and Howes, 1992). Extremely high C:P and N:P ratios of 3281 and 150, respectively, were observed in summer 2012 during the brown tide in the ML when scavenging of P by the dense bloom would have caused low P availability to the macroalgae.

#### 4.3. HABs as bio-indicators of sewage N enrichment

Measurement of stable C and N isotopes in primary producers and food webs has provided a powerful method to detect and track sewage pollution through aquatic ecosystems (Heaton, 1986; McClelland et al., 1997; Kendall et al., 2007; Risk et al., 2008; Olsen et al., 2010). The  $\delta^{13}$ C data showed an overall inverse correlation with  $\delta^{15}$ N, with lighter values reflecting lower salinity and lighter, terrestrial dissolved inorganic carbon (DIC) sources compared to more enriched values reflecting DIC from marine influenced sites in the IRL (Olsen et al., 2010). Macroalgae  $\delta^{15}$ N data collected IRLwide in this study showed a highly enriched overall mean value of  $\sim$ +6.3‰, a value remarkably similar to those reported for macroalgae in other urbanized, sewage-polluted coastal waters (Table 1). In Sarasota Bay, the Sarasota Bay Estuary Program has used their Comprehensive Conservation and Management Plan to implement a Wastewater Treatment and Reclamation Action Plan, which includes the consolidation and removal of wastewater treatment plants, removal of septic tanks, and requires N removal as part of the wastewater treatment process. As a result of their ongoing effort, nitrogen pollution has been reduced by 64% and is reflected in macroalgae  $\delta^{15}$ N values, which have been reported at the low end for sewage N ( $\sim$ 3.8%; Table 1), and the low abundance of drift algae and HABs in Sarasota Bay (Lapointe, unpublished data; SBEP, 2014).

The highly enriched  $\delta^{15}$ N values we observed in IRL macroalgae likely reflect the widespread reliance on OSTDS on the IRL watersheds. Although direct sewage discharges (outfalls) into the IRL were largely eliminated by 1996, the intended phase-out of OSTDS in highly urbanized areas on the IRL watersheds never happened as mandated in the IRL Act of 1990. Consequently, urbanization of the IRL watersheds in recent decades has continued to rely heavily on OSTDS, which we estimate to conservatively account for at least twice the N-loading as that from municipal sewage treatment plants (Table 4). Unlike the OSTDS, the partially treated municipal wastewater is not discharged into the IRL, but rather into deep injection wells or re-used for purposes such as irrigation (Table 4). Modeling studies indicate that OSTDS account for >90% of the N in groundwaters on IRL watersheds, and that this N ultimately discharges into the IRL (Horsley and Witten, 2000; GeoHydros, 2014). Even if only 50% of the estimated OSTDS N loads to groundwaters  $(2,120,013 \text{ kg N yr}^{-1}; \text{ Table 4})$  are currently reaching the IRL, this N load can more than account for the estimated total point and non-point source N loading to the IRL (893,053 kg N yr<sup>-1</sup>; Sigua and Tweedale, 2004). The discrepancies between FDOH reports for installed (299,612; FDOH, 2013) vs. active (72,441; FDOH unpublished geographic information systems data as of June 2012; GeoHydros, 2014) OSTDS and recent studies (212,100; GeoHydros, 2014) also highlights the uncertainty about the number, location, and loadings from OSTDS in Florida. Recent modeling studies show that OSTDS rank as the second largest source of N contamination to Florida's surface waters, just behind agriculture (Badruzzaman et al., 2012).

Nitrogen in OSTDS effluent is primarily in the form of ammonium (Bicki et al., 1984; Lapointe et al., 1990; Valiela et al., 1997) with  $\delta^{15}N$  values of ~+4–5‰ (Lapointe and Krupa, 1995; Hinkle et al., 2008; Katz et al., 2010). Ammonium containing the lighter <sup>14</sup>N isotope volatilizes preferentially, causing fractionation and enrichment of the remaining ammonium in the effluent

#### Table 4

Number of OSTDS and WWTPs in the five counties bordering the Indian River Lagoon. Specifically, the number of OSTDS and corresponding contributions of nitrogen to the IRL (GeoHydros, 2014) compared with number of WWTPs, flow rates, and calculated nitrogen contributions to obtain the percentage of OSTDS reliance for Volusia, Brevard, Indian River, St. Lucie, and Martin Counties (Englehardt et al., 2001; FDEP, 2014). Total % OSTDS reliance for all counties was calculated using an average of 2.45 people per OSTDS. Nitrogen input from WWTP disposal only includes facilities with a flow greater than 138,168 m<sup>3</sup>/yr and was calculated using total nitrogen concentrations found in secondary effluent (Englehardt et al., 2001).

County	# OSTDS	TN (kg/yr)	# WWTP (>138,168 m <sup>3</sup> /yr)	Flow (m <sup>3</sup> /yr)	TN (kg/yr)	Class I (deep) wells	# WWTP (<138,168 m <sup>3</sup> /yr)	Capacity (m³/yr) <sup>*</sup>	% OSTDS reliance
Volusia	77,705	784,595	16	45,899,326	275,990	0	84	1,951,896	38
Brevard	66,720	662,742	20	53,277,484	369,709	7	37	1,211,178	30
Indian River	28,889	276,322	6	10,597,466	43,923	1	7	4,861,432	48
St. Lucie	16,837	180,959	13	20,711,346	238,878	9	16	1,590,449	16
Martin	21,949	215,395	8	10,238,230	86,672	5	17	434,952	36
Total	212,100	2,120,013	63	140,723,853	1,015,172	22	161	10,049,908	32

Capacity of <138,168 m<sup>3</sup>/yr WWTP should not be interpreted as actual flow.

(Böhlke et al., 2006). If there is an adequate oxic unsaturated zone (vadose zone), the remaining ammonium can be nitrified to nitrate by bacteria; however, this microbial transformation is limited on IRL watersheds by the low elevations and high water tables that are indicative of "failing" OSTDS, which results in ammonium enrichment of groundwaters (Lapointe et al., 1990; Bicki and Brown, 1990; Lapointe and Krupa, 1995a,b). Considering the importance of SGD as a pathway for OSTDS-derived N transport into the IRL (Lapointe and Krupa, 1995a,b; Martin et al., 2007), the widespread use of OSTDS in generally poor soil conditions (IRLNEP, 1996) helps explain our observed dominance of ammonium rather than nitrate throughout the IRL. Macroalgae would preferentially assimilate NH4 compared to NO3 (D'Elia and DeBoer, 1978), a phenomenon that results in  $\delta^{15}$ N enrichment of these benthic matforming algae that are the "first responders" to SGD of wastewater N (Fig. 10). Nonetheless,  $\delta^{15}$ N-NO3 values from OSTDS are commonly heavier than  $\delta^{15}$ N-NH4 in septic discharges (Hinkle et al., 2008), which may reflect denitrification or reductive oxidation of ammonia and the subsequent loss of isotopically light N gas (Clark et al., 2008). However, denitrification is highly dependent on very specific environmental conditions (presence/ absence of suitable aerobic/anaerobic zones in effluent flow path, adequate carbon source(s); Kendall et al., 2007), which do not generally prevail in the sandy, low organic content, porous soils and shallow groundwaters on the IRL watersheds (Bicki et al., 1984; Woodward-Clyde, 1994; Horsley and Witten, 2000).

Fertilizers represent another potential N (and P) source in urban and agricultural watersheds of the IRL. Recent nutrient management policies adopted by counties and municipalities along the IRL have focused on moderating these inputs in a variety of urban settings. However, past studies suggest that fertilizer nutrient contributions may not be as important as commonly thought. Watershed-scale studies in Waquoit Bay, Cape Cod, MA showed that fertilizer inputs from urbanized watersheds, relative to OSTDS, were negligible, because landscape vegetation was highly efficient at nutrient uptake (Valiela et al., 1992); similar conclusions were reached for N-loading studies in Narragansett Bay, RI (Thornber et al., 2008). These conclusions are supported by experimental turf studies that reported minimal environmental effects of fertilizer N leaching into groundwaters and surface waters (Petrovic, 1990; Erickson et al., 2001), and that healthy, properly fertilized turf grass helps to prevent soil erosion, especially during active growth periods, when nutrient losses are negligible (Hochmuth et al., 2011). The fact that our  $\delta^{15}$ N values in IRL macroalgae so closely match  $\delta^{15}N$  values in Narragansett Bay and Waquoit Bay where N-loading is dominated by wastewater strongly points to wastewater, not fertilizers, as the primary N source in the IRL. The opposite can be seen in the low  $\delta^{15}$ N values (<3‰) of macroalgae in western Florida Bay that is downstream of agricultural watersheds where fertilizer and top soil N with relatively depleted  $\delta^{15}$ N values dominate the N supply (Lapointe et al., 2004).

In contrast to fertilizer N applications, N derived from OSTDS in urban landscapes enters the groundwater well below the thatch and root zone and therefore have relatively little opportunity for assimilation or uptake by vegetation. The groundwater-borne N is then transported downgradient with minimal loss of N via denitrification in the sandy, porous soils on IRL watersheds (Bicki et al., 1984). Considering the substantial N-loading from OSTDS on the IRL watersheds (Table 4; GeoHydros, 2014), we conclude that the  $\delta^{15}$ N enrichment in IRL macroalgae largely reflects cumulative OSTDS-derived N as it travels through the surficial groundwater and discharges into the IRL. This is a well-documented phenomenon in a wide variety of coastal settings (Lapointe et al., 1990; Valiela et al., 1992; Weiskel and Howes, 1992; Lapointe and Krupa, 1995a,b; Valiela et al., 1997; Reay, 2004; Mallin, 2013).

Highly enriched  $\delta^{15}$ N values were also recently observed in the Microcystis blooms sampled in the SLE during the summer of 2013. Unprecedented blooms of the Microcystis aeruginosa in the SLE formed following massive releases of fresh water from Lake Okeechobee and the C-44 watershed in 2005 and again in 2013. Blooms of M. aeruginosa have been increasing in freshwater lakes and rivers worldwide as a result of increasing N and P inputs from urban, agricultural, and industrial sources (Paerl and Otten, 2013). According to Parrish (2014), M. aeruginosa experiences optimal growth and abundance when the N:P ratio is <44:1, as seen in the SLE during the 2005 bloom when N:P ratios decreased to  $\sim$ 9:1 (Lapointe et al., 2012). Once transported into the SLE with the freshwater discharges, blooms of M. aeruginosa are likely fueled by nutrient inputs from the surrounding watershed, especially high P inputs from the C-23 and C-24 canals (Lapointe et al., 2012). The enriched  $\delta^{15}$ N value (+8.6‰) of the *M. aeruginosa* collected in August 2013 also points to wastewater N as a primary N source fueling the bloom. This highly urbanized estuary is locally impacted by thousands of OSTDS, many of which are "failing" and enrich tidal creeks and canals with high concentrations of ammonium, nitrate, and fecal bacteria (Lapointe et al., 2012). Interestingly, M. aeruginosa has a much higher affinity for ammonium than nitrate, further supporting the uptake of minimally-treated wastewater N in the SLE and the need for advanced wastewater treatment (N-removal) within the watershed (Parrish, 2014).

During the 2005 discharges from Lake Okeechobee to the Caloosahatchee River on Florida's west coast, similarly enriched  $\delta^{15}N$  values of +11.5‰ were measured in *M. aeruginosa* in the Caloosahatchee estuary and + 7.83‰ in red tides – *Karenia brevis* – in coastal waters off Sanibel Island (Yentsch et al., 2008). Like the SLE, the Caloosahatchee River receives considerable sewage N inputs from OSTDS, but also municipal surface water sewage outfalls (Lapointe and Bedford, 2007). In comparison, the  $\delta^{15}N$  value of ~+3.5‰ we measured in the superbloom in ML is at the low end of the wastewater N range, and could reflect minimal transformation/fractionation of sewage effluent as well as additional fertilizer and top soil N contributions that would have more depleted  $\delta^{15}N$  values, such as those reported for macroalgal blooms in western Florida Bay (~+2‰; Lapointe et al., 2004).

#### 4.4. Impacts of HABs on seagrasses and wildlife

With the high nitrogen concentrations and HABs during this study, also came the loss of seagrass coverage. This was reflected not only in our seagrass surveys, but also that of SJRWMD researchers. Between 2009 and 2012, over the course of the entire two-year study, seagrass coverage between Ponce and Ft. Pierce inlets decreased by about 60% with a loss of 47,000 acres (SIRWMD, 2013). Not only did overall seagrass coverage decrease within the IRL, but the northern extent of Thalassia testudinum also receded (SIRWMD, unpublished data). During the study period, T. testudinum was absent at the northern CIRL sites, including Sebastian Inlet, which was the previously known northern limit (Fig. 11; SJRWMD, unpublished data). In the ML, BR, and NIRL, where TDN values exceed 37 µM, there was no T. testudinum present. However, in the SIRL (Hobe Sound National Wildlife Refuge Nature Center to Jupiter Inlet), where T. testudinum abundance increases, the TDN values were well below this TN threshold (Fig. 11). The same pattern of decreased T. testudinum abundance with elevated TN concentrations was first reported for the Florida Keys, where only Halodule wrightii was present at transects in urbanized areas with dense OSTDS and eutrophic conditions (mean TN  $\sim$ 37  $\mu$ M); in comparison, *T. testudinum* was the dominant species of seagrass at other mesotrophic and oligotrophic locations (Lapointe et al., 1994). Although the exact TDN threshold where T. testudinum can grow may vary with location, higher TDN concentrations correlates with lower T. testudium and higher H. wrightii cover in a variety of Florida estuaries as well. In Florida Bay, for example, H. wrightii began to colonize and outcompete T. testudinum when TN levels increased (Fourgurean et al., 1995). This pattern is also illustrated in areas of Tampa Bay, where only sparse patches of *H. wrightii* are found in areas that receive high nitrogen loads (Hillsborough Bay) and T. testudinum dominates in areas with comparatively small nitrogen loads (Lower Tampa Bay; Coastal Resources Group Inc., 2009; Pribble et al., 2001; Yarbro and Carlson, 2013). Although there are many factors that can affect seagrass abundance and distribution, it appears that N levels can be a useful indicator of ecosystem health and species composition and that a threshold for sustained T. testudinum growth may exist. These dynamics between seagrass abundance, distribution and composition, and nutrient enrichment need to be better understood for proper management of seagrass habitats and the wildlife they support.

Following the unprecedented HABs and seagrass loss in 2011-2012, loss of food sources resulted in a trophic cascade and unusual mortality of wildlife in the northern IRL. Between 2012-2013, an unprecedented die-off of Atlantic bottlenose dolphins (Tursiops truncatus; ~100), West Indian manatees (Trichechus manatus Linnaeus; ~335), and brown pelicans (Pelecanus occidentalis Linnaeus; ~250) occurred in the NIRL and BR segments (Florida Fish and Wildlife Conservation Commission, unpublished data; Hubbs SeaWorld, unpublished data; Wines, 2013) prompting the National Oceanic and Atmospheric Administration (NOAA) to declare an Unusual Mortality Event in August 2013. IRL dolphins that stranded during this time were mal-nourished (NOAA, 2014). Petersen et al. (2013) links loss of seagrass to reduction in the number of pigfish (Orthopristis chrysoptera Linnaeus) and subsequently its predator the spotted seatrout (Cynoscion nebulosus Cuvier); the preferred food choice of the IRL dolphins. With the dramatic decline in seagrass cover manatees have had to supplement their diet with increased amounts of drift macroalgae, which continues to grow well in the shallow, hypereutrophic waters of the northern IRL segments. Ongoing research suggests that Gracilaria tikvahiae McLachlan, an abundant member of the drift algal community that is widely available as food to manatees, is producing N-containing cyanogenic glycosides (Peter Moeller et al., unpublished data) that are known to cause severe health problems, and even death, in humans (Seigler, 1991; Halstead and Haddock, 1992; Noguchi et al., 1994). In addition to the potential lethal effects of these macroalgal toxins, other more common HAB toxins, including brevetoxin, saxitoxin, and okadaic acid, are known to have sublethal effects on federally endangered manatees and green sea turtles in Florida (Capper et al., 2013). Because the eutrophication problem in the IRL appears to be driven largely by N from wastewater, there is a critical and urgent need for improved sewage collection and treatment - specifically involving Advanced Wastewater Treatment (AWT) and nutrient removal. Diversion of the N loading associated with sewage from the IRL would help restore the water quality conditions necessary for a healthy, seagrass-based ecosystem, a pre-requisite for sustaining populations of state and federally protected species.

#### 4.5. Economic impacts to IRL

As of 2007, the estimated annual economic value of the IRL was approximately \$3.7 billion (Hazen and Sawyer, 2008). The primary recreational uses of the lagoon are fin fishing (38%), swimming or wading (20%), and power boating (13%). Both commercial and recreational fishing activities are tied to the health of IRL seagrass beds. Based on this relationship, Hazen and Sawyer (2008) estimated IRL seagrass habitat (~72,400 acres in 2005) to be

worth approximately \$4,600 per acre per year or  $\sim$ \$227,000 per acre over the next 100 years. With a 60% loss of seagrass in the northern segments of the IRL (SJRWMD, 2013), the estimated economic impacts of the superbloom in 2011 and brown tides in 2012 and 2013 were substantial ( $\sim$ \$197 M loss/year).

#### 5. Conclusions

Although benthic macroalgal blooms have been common in seagrass habitats in the IRL for many decades (Benz et al., 1979), unprecedented phytoplankton blooms and seagrass die-off in the IRL during 2011–2012 indicates an ecosystem "tipping point" occurred during our study. The dominance of seagrass species (e.g., Halodule wrightii), with higher N tolerance (Fourgurean et al., 1995; Lapointe et al., 1994) combined with widespread macroalgal blooms and seagrass epiphyte loads in the NIRL is consistent with the classic model of nutrient enrichment and eutrophication in shallow, seagrass-dominated estuaries (Valiela et al., 1992; Bricker et al., 2007; Burkholder et al., 2007). The unprecedented phytoplankton "superbloom" we observed in 2011 followed a prolonged, 5-year drought and began following the onset of increased rainfall in spring 2011 (Fig. 2A). Such "first flush" stormwater events are well known to transport peak loads of nutrients from both urban stormwater runoff (Wanielista and Yousef, 1993) as well as OSTDS-contaminated groundwater discharges (Lapointe et al., 1990; Bicki and Brown, 1990). These rainfall-related increases in land-based nutrient inputs, especially P, help explain the high biomass phytoplankton blooms in the northern IRL segments in 2011-2012 where high TDN concentrations and TDN:TDP ratios resulted in strong P-limited conditions. The brown tide organism Aureoumbra lagunensis is well known to compete favorably with other phytoplankton under high DON concentrations (Gobler et al., 2013) and N:P ratios (Liu et al., 2001), an environmental condition that occurred in the ML and NIRL during our study.

Other researchers have suggested that hypersalinity in the ML as a result of the prolonged drought may have also contributed to the brown tide (Gobler et al., 2013). While hypersalinity (>40 psu) did occur in the Laguna Madre, TX during formation of the Aureoumbra lagunensis bloom, this organism has a broad tolerance to salinity and can grown between 10 and 90 psu (Buskey et al., 1998). The ML has had a long history of hypersaline conditions (>40) without the occurrence of brown tides, and hypersalinity has long been recognized as indicative of poor flushing and long residence times that are also conducive to bloom development (Briel et al., 1973; Buskey et al., 1998). Because freshwater inputs to the IRL are also the major source of N and P driving eutrophication, the issue of hypersalinity raises a note of caution to water managers considering a "quick fix." For example, similar phytoplankton blooms and seagrass die-off occurred in Florida Bay in the 1980s following diversion of N-rich agricultural runoff from Lake Okeechobee south to Everglades National Park and Florida Bay, which resulted in high N concentrations and N:P ratios similar to that in the northern IRL segments (Brand, 2002; Lapointe and Barile, 2004). Following a short drought in 1989–1990, Florida Bay researchers and water managers suggested that the phytoplankton blooms resulted from hypersalinity (baywide average  $\sim$ 41), which they claimed caused die-off of T. testudinum, subsequent internal nutrient enrichment and development of phytoplankton blooms. Water managers responded by further increasing flows of N-rich waters to both Shark River and Taylor Slough between 1991 and 1995, exacerbating nitrogen enrichment and phytoplankton blooms including red tide, *Karenia brevis*, causing more seagrass die-off, sponge die-off, regional-scale eutrophication, and unprecedented coral diseases and die-off in the Florida Keys National Marine Sanctuary (Lapointe and Barile, 2004). The decision to

increase freshwater flows into Florida Bay was a political decision, as no scientific evidence occurred at that time, or presently, that the hypersalinity in Florida Bay was either a primary or secondary factor in the die-off of *T. testudinum*. Hence, adding more freshwater to the ML and NIRL could similarly cause a worsening of the eutrophication problem in the IRL, as was the case in Florida Bay.

The common thread between eutrophication of the IRL and Florida Bay is the role of excess nitrogen in driving HABs and the loss of seagrasses and biodiversity, the latter being the biggest problem facing our planet (NRC, 2000; Rockström et al., 2009). In the case of the urbanized IRL, our results indicate that wastewater, and OSTDS in particular, is the major N source, compared to agricultural fertilizers and top soil N in Florida Bay (Brand, 2002; Glibert et al., 2004; Lapointe et al., 2004). These findings are consistent with recent modeling efforts showing that sewage is the primary source of N pollution in urban areas, compared to fertilizers in agricultural areas (McCrackin et al., 2013). Despite this knowledge, FDEP has emphasized WWTPs and stormwater in multiple TMDL and Basin Management Action Plans (BMAP) recently drafted for the IRL and several of its tributaries. Considering that point-source sewage inputs to the IRL were largely eliminated by the mid-1990s and that N loads from OSTDS are substantial (Table 4), we conclude that greater emphasis should instead be placed on quantifying non-point source nutrient pollution from the  $\sim$ 300,000 OSTDS in the IRL watershed; a major and growing N source to the system. This is particularly evident in the NIRL, ML, and BR where high TDN concentrations and TDN:TDP ratios reflect cumulative buildup and cycling of nitrogen-rich groundwater inputs. The high estimated N loadings from OSTDS suggest that previous nutrient loading models for the IRL have greatly underestimated the contribution of OSTDS as an N source to the IRL (GeoHydros, 2014). Thus, N loading from OSTDS and inadequate sewage treatment facilities should be at the forefront in planning documents, such as BMAPs, aimed at nutrient reduction to the IRL and its tributaries. Currently, any gains from nutrient-reducing BMAP projects are likely offset by the installation of additional OSTDS in newly developed areas.

#### Acknowledgements

The authors thank Ian MacLeod, Jeff Beal (Florida Fish and Wildlife Conservation Commission), Karen Holloway-Adkins and Doug Scheidt (Innovative Health Applications - Kennedy Space Center), and Gabrielle Barbarite (HBOI) for field and logistical support, James Nelson (HBOI) for boat support, and James Lyon and Mike Legare (U.S. Fish and Wildlife Service - Merritt Island National Wildlife Refuge) for permitting support. Lori Morris, Lauren Hall, Joel Steward, and Bob Chamberlain (St. Johns River Water Management District) provided information on seagrasses and photographs of macroalgae in the IRL. We also acknowledge Mike Bechtold (Florida Department of Environmental Protection) for providing access to wastewater data for the IRL. The study was funded through the Save Our Seas specialty license plate fund administered through the Harbor Branch Oceanographic Institute (HBOI) Foundation and supported through the work of Coastal Ocean Association of Science and Technology (COAST). This is contribution #1942 of the Harbor Branch Oceanographic Institute at Florida Atlantic University. [SS]

#### References

- Asplia, I., Agemian, H., Chau, A.S.Y., 1976. A semi-automated method for the determination of inorganic, organic, and total phosphate in sediments. Analyst 101, 187–197.
- Atkinson, M.J., Smith, S.V., 1983. C:N:P ratios of benthic marine plants. Limnol. Oceanogr. 28 (3), 568–574.

- Badruzzaman, M., Pinzon, J., Oppenheimer, J., Jacangelo, J.G., 2012. Sources of nutrients impacting surface waters in Florida: a review. J. Environ. Manag. 109, 80–92.
- Barile, P.J., 2004. Evidence of anthropogenic nitrogen enrichment of the littoral waters of east central Florida. J. Coast. Res. 20 (4), 1237–1245.
- Bateman, A.S., Kelly, S.D., 2007. Fertilizer nitrogen isotope signatures. Isotopes Environ. Health Stud. 43 (3), 237–247.
- Belanger, T.V., Price, T.L., Heck, Howell, H., 2007. Submarine groundwater discharge in the Indian River Lagoon Florida. How important is it? Fla. Sci. 70 (4), 344–362.
- Benz, M.C., Eiseman, N.J., Gallaher, E.E., 1979. Seasonal occurrence and variation in standing crop of a drift algal community in the Indian River, Florida. Bot. Mar. 22, 413–420.
- Bicki, T.J., Brown, R.B., 1990. On-site sewage disposal: the importance of the wet season water table. J. Environ. Health 52 (5), 277–279.
- Bicki, T.J., Brown, R.B., Collins, M.E., Mansell, R.S., Rothwell, D.F., 1984. Impact of on-site sewage disposal systems on surface and ground water quality. Report to Florida Department of Health and Rehabilitative Services under contract number LC170.
- Böhlke, J.K., Smith, R.L., Miller, D.N., 2006. Ammonium transport and reaction in contaminated groundwater: application of isotope fractionation studies. Water Resour. Res. 42 (5), 2006.
- Bossart, G.D., Romano, T.A., Peden-Adams, M.M., Schaefer, A.M., McCulloch, S., Goldstein, J.D., Rice, C.D., Fair, P.A., Cray, C., Reif, J.S., 2014. Clinicoimmunopathologic findings in Atlantic bottlenose dolphins *Turisops truncatus* with positive *Chlamydiaceae* antibody titers. Dis. Aquat. Organ. 108, 71–81.
- Brand, L.E., 2002. The transport of terrestrial nutrients to South Florida coastal waters. In: Porter, J.W., Porter, K.G. (Eds.), The Everglades, Florida Bay, and Coral Reefs of the Florida Keys. CRC Press, Boca Raton, FL.
- Brezonik, P.L., 1972. Nitrogen: sources and transformations in natural waters. In: Allen, H.E., Kramer, J.R. (Eds.), Nutrients in Natural Waters. John Wiley & Sons, New York, pp. 1–50.
- Bricker, S., Longstaff, B., Dennison, W., Jones, A., Boicourt, K., Wicks, C., Woerner, J., 2007. Effects of Nutrient Enrichment in the Nation's Estuaries: A Decade of Change. National Centers for Coastal Ocean Science, Silver Spring, MD.
- Briel, L.I., Fyler, J.M., Laffey, M.Y., Paxton, J.T., Stephens, F.C., 1973. Water quality studies of the Indian River Lagoon. Indian River Study, Annual Report 1973– 1974, vol. 1Harbor Branch Consortium, Fort Pierce, FL, pp. 56–90 (Chapter 4).
- Bronk, D.A., Glibert, P.M., Malone, T.C., Banahan, S., Sahlsten, E., 1998. Inorganic and organic nitrogen cycling in Chesapeake Bay: autotrophic versus heterotrophic processes and relationships to carbon flux. Aquat. Microb. Ecol. 15, 177–189.
- Burkholder, J.M., Tomasko, D.A., Touchette, B.W., 2007. Seagrasses and eutrophication. J. Exp. Mar. Biol. Ecol. 350, 46–72.
- Buskey, E.J., Wysor, B., Hyatt, C., 1998. The role of hypersalinity in the persistence of the Texas 'brown tide' in the Laguna Madre. J. Plankton Res. 20 (8), 1553–1565.
- Buskey, E.J., Liu, H.B., Collumb, C., Bersano, J.G.F., 2001. The decline and recovery of a persistent Texas brown tide algal bloom in the Laguna Madre (Texas, USA). Estuaries 24, 337–346.
- Capper, A., Flewelling, L.J., Arthur, K., 2013. Dietary exposure to harmful algal bloom (HAB) toxins in the endangered manatee (*Trichechus manatus latirostris*) and green sea turtle (*Chelonia mydas*) in Florida. USA. Harmful Algae 28, 1–9.
- Clark, I., Timlin, R., Bourbonnais, A., Jones, K., Lafleur, D., Wickens, K., 2008. Origin and fate of industrial ammonium in anoxic ground water-<sup>15</sup>N evidence for anaerobic oxidation (Anammox). Ground Water Monit. Remediation 28 (3), 73–82.
- Coastal Resources Group Inc., 2009. "Knowledge-based" seagrass maps for Tampa Bay, Florida. Prepared for Tampa Bay Estuary Program 8 pp.
- Costanzo, S.D., O'Donohue, M.J., Dennison, W.C., Loneragan, N.R., Thomas, M., 2001. A new approach for detecting and mapping sewage impacts. Mar. Pollut. Bull. 42 (2), 149–156.
- D'Elia, C.F., DeBoer, J.A., 1978. Nutritional studies of two red algae: 2. Kinetics of ammonium and nitrate uptake. J. Phycol. 14, 266–272.
- D'Elia, C.F., Connor, E.E., Kaumeyer, N.L., Keefe, C.W., Wood, K.V., Zimmerman, C.F., 1997. Nutrient analytical services: standard operating procedures. Technical Report Series No. 158-97.
- Dolman, A.M., Rücker, J., Pick, F.R., Fastner, J., Rohrlack, T., Mischke, U., Wiedner, C., 2012. Cyanobacteria and cyanotoxins: the influence of nitrogen versus phosphorus. PLoS ONE 7 (6), e38757, http://dx.doi.org/10.1371/journal.pone.0038757.
- Donald, D.B., Bogard, M.J., Finlay, K., Leavitt, P.R., 2011. Comparative effects of urea, ammonium, and nitrate on phytoplankton abundance, community composition, and toxicity in hypereutrophic freshwaters. Limnol. Oceanogr. 56 (6), 2161–2175.
- Duncan, B.W., Larson, V.L., Schmalzer, P.A., 2004. Historical and recent landscape change in the north Indian River Lagoon Watershed, Florida, USA. Nat. Areas J. 24, 198–215.
- Englehardt, J.D., Amy, V.P., Bloetscher, F., Chin, D.A., Fleming, L.E., Gokgoz, S., Rose, J.B., Solo-Gabriele, H., Tchobanoglous, G., 2001. Comparative assessment of human and ecological impacts from municipal wastewater disposal methods in Southeast Florida. Prepared for Florida Water Environment Association Utility Council July 12.
- Erickson, J.E., Cisar, J.L., Volin, J.C., Snyder, G.H., 2001. Comparing nitrogen runoff and leaching between newly established St Augustine grass turf and an alternative residential landscape. Crop Sci. 41, 1889–1895.
- FDEP (Florida Department of Environmental Protection), 2014. 2013 Water Reuse Inventory. Florida Department of Environmental Protection, Tallahassee, Florida.

- FDEP (Florida Department of Environmental Protection), 2008. Water Quality Assessment Report- Indian River Lagoon, Division of Environmental Assessment and Restoration, http://www.dep.state.fl.us/water/basin411/indianriver/ assessment.htm.
- FDOH (Florida Department of Health), 2013. Onsite Sewage Treatment and Disposal Systems Installed in Florida, http://www.floridahealth.gov/environmentalhealth/onsite-sewage/\_documents/new-installations.pdf.
- Fourqurean, J.W., Powell, G.V.N., Kenworthy, W.J., Zeiman, J.C., 1995. The effects of long-term manipulation of nutrient supply on competition between the seagrasses *Thalassia testudinum* and *Halodule wrightii* in Florida Bay. Oikos 72, 349–385.
- France, R., Holmquist, J., Chandler, M., Cattaneo, A., 1998. δ<sup>15</sup>N evidence for nitrogen fixation associated with macroalgae from a seagrass-mangrove-coral reef system. Mar. Ecol. Prog. Ser. 167, 297–299.
- Fujimoto, N., Sudo, R., Sugiura, N., Inamori, Y., 1997. Nutrient-limited growth of *Microcystis aeruginosa* and *Phormidium tenue* and competition under various N:P supply ratios and temperatures. Limnol. Oceanogr. 42 (2), 250–256.
- GeoHydros, 2014. Contributions of Total Nitrogen from OSTDS to the Indian River Lagoon and the Wakulla-St. Marks River Drainage Basins, Florida. Technical Memorandum, Prepared for Coastal Ocean Association of Science & Technology. April 25, 2014.
- Gilbert, S., Clark, K., 1981. Seasonal variation in standing crop of the seagrass Syringodium filiforme and associated macrophytes in the northern Indian River, FL. Estuaries 4, 223–225.
- Glibert, P.M., Heil, C.A., Hollander, D.J., Revilla, M., Hoare, A., Alexander, J., Murasko, S., 2004. Evidence for dissolved organic nitrogen and phosphorus uptake during a cyanobacterial bloom in Florida Bay. Mar. Ecol. Prog. Ser. 280, 73–83.
- Gobler, C.J., Sunda, W.G., 2012. Ecosystem disruptive algal blooms of the brown tide species Aureococcus anophagefferens and Aureoumbra lagunensis. Harmful Algae 14, 36–45.
- Gobler, C.J., Koch, F., Kang, Y., Berry, D.L., Tang, Y.Z., Lasi, M., Walters, L., Hall, L., Miller, J.D., 2013. Expansion of harmful brown tides caused by the pelagophyte *Aureoumbra lagunensis* De Yoe et Stockwell, to the US east coast. Harmful Algae 27, 29–41.
- Hall, L., Morris, L., Virnstein, R., Carter, E., 2001. Seagrass and drift algae biomass estimates for the Indian River Lagoon, FL. St Johns River Water Management District (Unpublished Technical Report).
- Halstead, B.W., Haddock, R.L., 1992. A fatal outbreak of poisoning from the ingestion of red seaweed *Gracilaria tsudae* in Guam – a review of the oral marine biotoxicity problem. J. Nat. Toxins 1, 87–115.
- Hanisak, M.D., 1990. The use of *Gracilaria tikvahiae* as a model system to understand the nitrogen nutrition of culture seaweeds. In: Lindstrom, S.C., Gabrielson, P.W. (Eds.), Thirteenth International Seaweed Symposium, Developments in Hydrobiology, 58. pp. 79–87.
- Hardison, D.R., Sunda, W.G., Shea, D., Litaker, R.W., 2013. Increased toxicity of *Karenia brevis* during phosphate limited growth: ecological and evolutionary implications. PLoS ONE 8 (3), e58545, http://dx.doi.org/10.1371/journal.pone.0058545.
- Hazen and Sawyer, P.C., Milian, Swain & Associates Inc., 2008. Indian River Lagoon economic assessment and analysis update. Final report for the Indian River Lagoon National Estuary Program. August 18, 2008 pages by chapter not consecutive.
- Heaton, T.H.E., 1986. Isotopic studies of nitrogen pollution in the hydrosphere and atmosphere: a review. Chem. Geol. 5, 87–102.
- Hinkle, S.R., Böhlke, J.K., Fisher, L.H., 2008. Mass balance and isotope effects during nitrogen transport through septic tank systems with packed-bed (sand) filters. Sci. Total Environ. 407, 324–332.
- Hirama, S., Ehrhart, L.M., Rea, L.D., Kiltie, R.A., 2014. Relating fibropapilloma tumor severity to blood parameters in green turtles *Chelonia mydas*. Dis. Aquat. Organ. 111, 61–68.
- Hochmuth, G., Nell, T., Sartain, J., Unruh, J.B., Martinez, C., Trenholm, L., Cisar, J., 2011. Urban water quality and fertilizer ordinances: Avoiding unintended consequences: A review of the scientific literature. Soil and Water Science Department, UF/IFAS Extension, Publication # SL 283 25 pp.
- Horsley and Witten Inc., 2000. On-site sewage disposal systems pollutant loading evaluation. Test and validation of Indian River Lagoon nitrogen model. Indian River Lagoon Program, Melbourne, FL 43 pp.
- Howarth, R.W., Marino, R., 2006. Nitrogen as the limiting nutrient for eutrophication in coastal marine ecosystems: evolving views over three decades. Limnol. Oceanogr. 51 (Part 2 (1)), 364–376.
- IRLNEP (Indian River Lagoon National Estuary Program), 1996. Indian River Lagoon Comprehensive Conservation and Management Plan. Indian River Lagoon National Estuary Program, Melbourne, FL 357 pp.
- IRLNEP (Indian River Lagoon National Estuary Program), 2008. Indian River Lagoon Comprehensive Conservation and Management Plan Update. Indian River Lagoon National Estuary Program, Palm Bay, FL 120 pp.
- Katz, B.G., Griffin, D.W., McMahon, P.B., Harden, H.S., Wade, E., Hicks, R.W., Chanton, J.P., 2010. Fate of effluent-borne contaminants beneath septic tank drainfield overlying a karst aquifer. J. Environ. Qual. 39, 1181–1195.
- Kendall, C., Elliott, E.M., Wankel, S.D., 2007. Tracing anthropogenic inputs of nitrogen to ecosystems. In: Michener, R.H., Lajtha, K. (Eds.), Stable Isotopes in Ecology and Environmental Science. 2nd ed. Blackwell Publishing, (Chapter 12), pp. 375–449.
- Kim, Y., Engel, B.A., Lim, K.J., Larson, V., Duncan, B., 2002. Runoff impacts of land-use change in Indian River Lagoon watershed. J. Hydrol. Eng. 7 (3), 245–251.

- Kroening, S., 2007. Assessment of water-quality monitoring and a proposed waterquality monitoring network for the Mosquito Lagoon basin East-Central Florida. U.S. Department of the Interior & U.S. Geological Survey.
- Lapointe, B.E., 1985. Strategies for pulsed nutrient supply to *Gracilaria* cultures in the Florida Keys: interactions between concentration and frequency of nutrient pulses. J. Exp. Mar. Biol. Ecol. 93 (3), 211–222.
- Lapointe, B.E., 2013. Ecology and nutrition of macroalgal blooms in Sarasota Bay, Florida. Final Report. Prepared for Sarasota Bay Estuary Program 7 pp.
- Lapointe, B.E., Ryther, J.H., 1979. Effects of nitrogen and seawater flow-rate on the growth and biochemical composition of *Gracilaria foliifera* var. angustissima in mass outdoor culture. Bot. Marina 22 (8), 529–537.
- Lapointe, B.E., O'Connell, J., 1989. Nutrient-enhanced growth of *Cladophora prolifera* in Harrington Sound Bermuda: eutrophication of a confined, phosphoruslimited marine ecosystem. Estuar. Coast. Shelf Sci. 28, 347–360.
- Lapointe, B.E., Krupa, S., 1995a. Jupiter Creek septic tank/water quality investigation. Final Report. Loxahatchee River Environmental Control District, Jupiter, FL 96 pp.
- Lapointe, B.E., Krupa, S., 1995b. Tequesta Peninsula septic tank/water quality investigation. Final Report. Loxahatchee River Environmental Control District, Jupiter, FL 87 pp.
- Lapointe, B.E., Barile, P.J., 2004. Comment on J.C. Zieman, J.W. Fourqurean, and T.A. Frankovich. 1999. Seagrass die-off in Florida Bay: Long-term trends in abundance and growth of Turtle Grass, *Thalassia testudinum*. Estuaries 22:460–470. Estuaries 27 (1), 157–158.
- Lapointe, B.E., Bedford, B.J., 2007. Drift rhodophyte blooms emerge in Lee County, Florida, USA: evidence of escalating coastal eutrophication. Harmful Algae 6, 421–437.
- Lapointe, B.E., Barile, P.J., Littler, M.M., Littler, D.S., 2005a. Macroalgal blooms on southeast Florida coral reefs: II. Cross-shelf discrimination of nitrogen sources indicates widespread assimilation of sewage nitrogen. Harmful Algae 4 (6), 1106–1112.
- Lapointe, B.E., Dawes, C.J., Tenore, K.R., 1984. Interactions between light and temperature on the physiological ecology of *Gracilaria tikvahiae* (Gigartinales: Rhodophyta). II: Nitrate uptake and levels of pigments and chemical constituents. Mar. Biol. 80 (2), 171–178.
- Lapointe, B.E., Littler, M.M., Littler, D.S., 1992. Nutrient availability to marine macroalgae in siliciclastic versus carbonate-rich coastal waters. Estuaries 15, 75–82.
- Lapointe, B.E., Matzie, W.R., Barile, P.J., 2004. Anthropogenic nutrient enrichment of seagrass and coral reef communities in the lower Florida keys: discrimination of local versus regional nitrogen sources. J. Exp. Mar. Biol. Ecol. 308 (1), 23–58.
- Lapointe, B.E., O'Connell, J.D., Garrett, G.S., 1990. Nutrient couplings between onsite sewage disposal systems, groundwaters, and nearshore surface waters of the Florida Keys. Biogeochemistry 10, 289–307.
- Lapointe, B.E., Tomasko, D.A., Matzie, W.R., 1994. Eutrophication and trophic state classification of seagrass communities in the Florida Keys. Bull. Mar. Sci. 54 (3), 696–717.
- Lapointe, B.E., Barile, P.J., Wynne, M.J., Yentsch, C.S., 2005b. Reciprocal Caulerpa invasion: mediterranean native *Caulerpa ollivieri* in the Bahamas supported by human nitrogen enrichment. Aquat. Invaders 16 (2), 1–4.
- Lapointe, B.E., Langton, R., Bedford, B.J., Potts, A.C., Day, O., Hu, C., 2010. Land-based nutrient enrichment of the Buccoo Reef Complex and fringing coral reefs of Tobago, West Indies. Mar. Pollut. Bull. 60 (3), 334–343.
   Lapointe, B.E., Thacker, K., Hanson, C., Getten, L., 2011. Sewage pollution in Negril,
- Lapointe, B.E., Thacker, K., Hanson, C., Getten, L., 2011. Sewage pollution in Negril, Jamaica: effects on nutrition and ecology of coral reef macroalgae. Chin. J. Oceanol. Limn. 29 (4), 775–789.
- Lapointe, B.E., Herren, L.W., Bedford, B.J., 2012. Effects of hurricanes, land-use, and water management on nutrient and microbial pollution: St Lucie Estuary, southeast Florida. J. Coast. Res. 28 (6), 1345–1361.
- Lavrentyev, P.J., Bootsma, H.A., Johengen, T.H., Cavaletto, J.F., Gardner, W.S., 1998. Microbial plankton response to resource limitation: insights from the community structure and seston stoichiometry in Florida Bay, USA. Mar. Ecol. Prog. Ser. 165, 45–57.
- Littler, D.S., Littler, M.M., Hanisak, M.D., 2008. Submersed Plants of the Indian River Lagoon. Offshore Graphics, Inc., Washington, DC 286 pp.
- Liu, H., Laws, E.A., Villareal, T.A., Buskey, E.J., 2001. Nutrient-limited growth of Aureoumbra lagunensis (Pelagophyceae), with implications for its capabilities to outgrow other phytoplankton species in phosphate-limited environments. J. Phycol. 37, 500–508.
- Lu, Z., Gan, J., 2014. Controls of seasonal variability of phytoplankton blooms in the Pearl River Estuary. Deep-Sea Res., http://dx.doi.org/10.1016/j.dsr2. 2013.12.011 (in press).
- Lucas, L.V., Koseff, J.R., Monismith, S.G., Cloern, J.E., Thompson, J.K., 1999. Processes governing phytoplankton blooms in estuaries: II. The role of horizontal transport. Mar. Ecol. Prog. Ser. 187, 17–30.
- Mallin, M.A., 2013. Septic Systems in the Coastal Environment: Multiple Water Quality Problems in Many Areas. Monitoring Water Quality. Elsevier (Chapter 4).
- Martin, J.B., Cable, J.E., Smith, C., Roy, M., Cherrier, J., 2007. Magnitudes of submarine groundwater discharge from marine and terrestrial sources: Indian River Lagoon, Florida. Water Resour. Res. 43 (5), W05440, http://dx.doi.org/10.1029/ 2006WR005266.
- McClelland, J.W., Valiela, I., 1998. Changes in food web structure under the influence of increased anthropogenic nitrogen inputs to estuaries. Mar. Ecol. Prog. Ser. 168, 259–271.

- McClelland, J.W., Valiela, I., Michener, R., 1997. Nitrogen-stable isotope signatures in estuarine food webs: a record of increasing urbanization in coastal watersheds. Limnol. Oceanogr. 42 (5), 93–937.
- McCrackin, M.L., Harrison, J.A., Compton, J.E., 2013. A comparison of NEWS and SPARROW models to understand sources of nitrogen delivered to US coastal areas. Biogeochemistry 114, 281–297.
- McGlathery, KJ, 1995. Nutrient and grazing influences on a subtropical seagrass community. Mar. Ecol. Prog. Ser. 122, 239–252.
- MEA (Marine Ecosystem Assessment), 2005. Marine and Coastal Ecosystems and Human Well-Being: Synthesis. Prepared by UNEP World Conservation Monitoring Centre, Cambridge, UK. Island Press, Washington, DC 137 pp.
- Morris, L.J., Hall, L.M., Virnstein, R.W., 2001. Field guide for fixed seagrass transect monitoring in the Indian River Lagoon. Technical Memorandum. St. Johns River Water Management District, Palatka, FL 28 pp.
- Nixon, S.W., 1995. Coastal marine eutrophication: a definition, social causes, and future consequences. OPHELIA 41, 199–219.
- NOAA (National Oceanic and Atmospheric Administration), National Marine Fisheries Service, 2014. 2013 bottlenose dolphin unusual mortality event in Florida, http://www.nmfs.noaa.gov/pr/health/mmume/floridadolphins2013.html (accessed 17.10.14).
- Noguchi, T., Matsui, T., Miyazawa, K., Asakawa, M., luima, N., Shida, Y., Fuse, M., Hosaka, Y., Kirigaya, C., Watabe, K., Usui, S., Fukagawa, A., 1994. Poisoning by the red alga 'Ogonori' (*Gracilaria verrucosa*) on the Nojima Coast, Yokohama, Kanagawa Prefecture, Japan. Toxicon 32, 1533–1538.
- NRC (National Research Council), 1993. Managing Wastewater in Coastal Urban Areas. National Academy Press, Washington, DC 477 pp.
- NRC (National Research Council), 2000. Clean Coastal Waters: Understanding and Reducing the Effects of Nutrient Pollution. National Research Council, National Academy Press, Washington, DC 391 pp.
- Olsen, Y.S., Fox, S.E., Kinney, E.L., Teichberg, M., Valiela, I., 2010. Differences in urbanization and degree of marine influence are reflected in δC and δN of producers and consumers in seagrass habitats of Puerto Rico. Mar. Environ. Res. 69 (3), 198–206.
- Onuf, C.P., 1996. Seagrass responses to long-term light reduction by brown tide in upper Laguna Madre Texas: distribution and biomass patterns. Mar. Ecol. Prog. Ser. 138, 219–231.
- Paerl, H.W., Fogel, M.L., 1994. Isotopic characterization of atmospheric nitrogen inputs as sources of enhanced primary production in coastal Atlantic Ocean waters. Mar. Biol. 119 (4), 635–645.
- Paerl, H.W., Fulton III, R.S., 2006. Ecology of harmful cyanobacteria Ecology of Harmful Algae. Ecological Studies, vol. 189, pp. 95–109 (Chapter 8).
- Paerl, H.W., Otten, T.G., 2013. Harmful cyanobacterial blooms: causes, consequences, and controls. Microb. Ecol. 65 (4), 995–1010.
- Parrish, J., 2014. The Role of Nitrogen and Phosphorus in the Growth, Toxicity, and Distribution of the Toxic Cyanobacteria, *Microcystis aeruginosa*. (Master's thesis)University of San Francisco 57 pp.
- Petersen, K., Ramirez, D., Anderson, G., Mazzoil, M., Morris, L., O'Corry-Crowe, G., Hanisak, M.D., Gilmore Jr., R.G., 2013. Seagrass and dolphins: What can they tell us about the health of the Indian River Lagoon? Indian River Lagoon Symposium 2013: The Health of the Lagoon. Johnson Education Center HBOI-FAU, Fort Pierce, FL February 7–8, 2013.
- Petrovic, A.M., 1990. The fate of nitrogenous fertilizers applied to turfgrass. J. Environ. Qual. 19, 1–14.
- Phlips, E.J., Badylak, S., Christman, Lasi, M., 2010. Climatic trends and temporal patterns of phytoplankton composition, abundance, and succession in the Indian River Lagoon, Florida, USA. Estuar. Coasts 33, 498–512.
- Phlips, E.J., Badylak, S., Hart, J., Haunert, D., Lockwood, J., O'Donnell, K., Sun, D., Viveros, P., Yilmaz, M., 2012. Climatic influences on autochthonous and allochthonous phytoplankton blooms in a subtropical estuary, St. Lucie Estuary, Florida, USA. Estuar. Coasts 35, 335–352.
- Phlips, E.J., Badylak, S., Lasi, M.A., Chamberlain, R., Green, W.C., Hall, L.M., Hart, J.A., Lockwood, J.C., Miller, J.D., Morris, L.J., Steward, J.S., 2014. From red tides to green and brown tides: bloom dynamics in a restricted subtropical lagoon under shifting climatic conditions. Estuar. Coasts, http://dx.doi.org/10.1007/ s12237-014-9874-6.
- Phlips, E.J., Badylak, S., Christman, M., Wolny, J., Brame, J., Garland, J., Hall, L., Hart, J., Landsberg, J., Lasi, M., Lockwood, J., Paperno, R., Scheidt, D., Staples, A., Steidinger, K., 2011. Scales of temporal and spatial variability in the distribution of harmful algae species in the Indian River Lagoon, Florida, USA. Harmful Algae 10 (3), 277–290.
- Pribble, R., Janicki, A., Zarbock, H., Janicki, S., Winowitch, M., 2001. Estimates of total nitrogen, total phosphorus, total suspended solids, and biochemical oxygen demand loadings to Tampa Bay, Florida: 1995–1998. Prepared for Tampa Bay Estuary Program by Janicki Environmental, Inc. 227 pp.
- Reay, W.G., 2004. Septic tank impacts on ground water quality and nearshore sediment nutrient flux. Ground Water 42 (7), 1079–1089.
- Rhudy, K.B., Sharma, V.K., Lehman, R.L., McKee, D.A., 1999. Seasonal variability of the Texas 'Brown Tide' (*Aureoumbra lagunensis*) in relation to environmental parameters. Estuar. Coast. Shelf Sci. 48 (5), 565–574.
- Richardson, B., 2009. Physiological characteristics and competitive strategies of bloom-forming cyanobacteria and diatoms in Florida Bay. Bull. Mar. Sci. 38, 19–36.
- Riegl, B.M., Moyer, R.P., Morris, L.J., Virnstein, R.W., Purkis, S.J., 2005. Distribution and seasonal biomass of drift macroalgae in the Indian River Lagoon (Florida, USA) estimated with acoustic seafloor classification (QTCView Echoplus). J. Exp. Mar. Biol. Ecol. 326, 89–104.

- Risk, M.J., Lapointe, B.E., Sherwood, O.A., Bedford, B.J., 2008. The use of  $\delta^{15}$ N in assessing sewage stress on coral reefs. Mar. Pollut. Bull. 58 (6), 793–802.
- Rockström, J., Steffen, W., Noone, K., Persson, Å., Chapin III, F.S., Lambin, E.F., Lenton, T.M., Scheffer, M., Folke, C., Schellnhuber, H.J., Nykvist, B., de Wit, C.A., Hughes, T., van der Leeuw, S., Rodhe, H., Sörlin, S., Snyder, P.K., Costanza, R., Svedin, U., Falkenmark, M., Karlberg, L., Corell, R.W., Fabry, V.J., Hansen, J., Walker, B., Liverman, D., Richardson, K., Crutzen, P., Foley, J.A., 2009. A safe operating space for humanity. Nature 461, 472–475.
- SBEP (Sarasota Bay Estuary Program), 2014. Sarasota Bay comprehensive conservation and management plan update & state of the bay report. SBEP, Sarasota, FL 79 pp.
- Schaefer, A.M., Goldstein, J.D., Reif, J.S., Fair, P.A., Bossart, G.D., 2009. Antibioticresistant organisms cultured from Atlantic Bottlenose Dolphins (*Tursiops truncatus*) inhabiting estuarine waters of Charleston, SC and Indian River Lagoon, FL. EcoHealth 6 (1), 33–41.
- Schaefer, A.M., Bossart, G.D., Mazzoil, M., Fair, P.A., Reif, J.S., 2011. Risk factors for colonization of *E. coli* in Atlantic Bottlenose Dolphins (*Tursiops truncatus*) in the Indian River Lagoon, Florida. Environ. Public Health, http://dx.doi.org/10.1155/ 2011/59707, Article ID 597073, 8 pp.
- Seigler, D.S., 1991. Cyanide and cyanogenic glycosides. In: Rosenthal, G.A., Berenbaum, M.R. (Eds.), Herbivores: Their Interactions with Secondary Plant Metabolites, Vol. 1: The Chemical Participants, 2nd ed. Academic Press, Inc., San Diego, CA, 468 pp. (Chapter 2).
- Sigua, G.C., Tweedale, W.A., 2003. Watershed scale assessment of nitrogen and phosphorus loadings in the Indian River Lagoon basin Florida. J. Environ. Manag. 67 (4), 363–372.
- Sigua, G.C., Tweedale, W.A., 2004. Assessing redesigned effectiveness of the water quality monitoring program in the Indian River Lagoon, Florida. Aquat. Conserv. 14, 49–64.
- Sigua, G.C., Steward, J.S., Tweedale, W.A., 2000. Water quality monitoring and biological integrity assessment in the Indian River Lagoon, Florida: status, trends, and loadings (1988–1994). Environ. Manage. 25 (2), 199–209.
- SJRWMD (St. Johns River Water Management District), 2007. Indian River Lagoon: An Introduction to a National Treasure. St. Johns River Water Management District Indian River Lagoon National Estuary Program, http://www.sjrwmd. com/itsyourlagoon/pdfs/IRL\_Natural\_Treasure\_book.pdf.
- SJRWMD (St. Johns River Water Management District), 2013. The Indian River Lagoon: An estuary of national significance Floridaswater.com, http:// floridaswater.com/itsyourlagoon/.
- SJRWMD (St. Johns River Water Management District), Bethune-Cookman University, Florida Atlantic University-Harbor Branch Oceanographic Institution, Florida Fish and Wildlife Conservation Commission, Florida Institute of Technology, Nova Southeastern University, Smithsonian Marine Station at Fort Pierce, University of Florida, Seagrass Ecosystems Analysts, 2012. Indian River Lagoon 2011 Superbloom Plan of Investigation. Technical Report 26 pp.
- Smith, N.P., 1993. Tidal and non-tidal flushing of Florida's Indian River Lagoon. Estuaries 16 (4), 739-746.
- Steward, J.S., VanArman, J.A., 1987. Indian River Lagoon Joint Reconnaissance Report. St. Johns River Water Management District and South Florida Water Management District, Final Report to Department of Environmental Regulation and OCRM/NOAA Contract No. CM-137.
- Steward, J.S., Virnstein, R.W., Morris, L.J., Lowe, E.F., 2005. Setting seagrass depth, coverage, and light targets for the Indian River Lagoon System, Florida. Estuaries 28 (6), 923–935.
- Swain, H.M., Breininger, D.R., Busby, D.S., Clark, K.B., Cook, S.B., Day, R.A., De Freese, D.E., Gilmore, R.G., Hart, A.W., Hinkle, C.R., McArdle, D.A., Mikkelsen, P.M., Nelson, W.G., Zahorcak, A.J., 1995. Indian River Lagoon conference – introduction. Bull. Mar. Sci. 57 (1), 1–7.
- Terry, K.L., Laws, E.A., Burns, D.J., 1985. Growth rate and variation in the N:P requirement ratio of phytoplankton. J. Phycol. 21 (2), 323–329.
- Thornber, C.S., DiMilla, P., Nixon, S.W., McKinney, R.A., 2008. Natural and anthropogenic nitrogen uptake by bloom-forming macroalgae. Mar. Pollut. Bull. 56, 261–269.
- Tomasko, D.A., Lapointe, B.E., 1991. Productivity and biomass of *Thalassia testudinum* as related to water column nutrient availability and epiphyte levels: field observations and experimental studies. Mar. Ecol. Prog. Ser. 75, 9–17.
- Tyler, A.C., McGlathery, K.J., Anderson, I.C., 2001. Macroalgae mediation of dissolved organic nitrogen fluxes in a temperate coastal lagoon. Estuar. Coast. Shelf Sci. 53 (2), 155–168.
- US Census Bureau, 2014. Annual Estimates of the Resident Population: April 1, 2010 to July 1, 2013. http://factfinder.census.gov/faces/tableservices/jsf/pages/productview.xhtml?src=bkmk.
- US EPA (U.S. Environmental Protection Agency, 2002. Environmental Protection Agency Onsite wastewater treatment systems manual. EPA/625/R-00/008. Office of Water, Office of Research and Development.
- Valiela, I., Foreman, K., LaMontagne, M., Hersh, D., Costa, J., Peckol, P., DeMeo-Anreson, B., D'Avanzo, C., Babione, M., Sham, C.H., Brawley, J., Lajtha, K., 1992. Couplings of watersheds and coastal waters: sources and consequences of nutrient enrichment in Waquoit Bay Massachusetts. Estuaries 15 (4), 443–457.
- Valiela, I., McClelland, J., Hauxwell, J., Behr, P.J., Hersh, D., Foreman, K., 1997. Macroalgal blooms in shallow estuaries: controls and ecophysiological and ecosystem consequences. Limnol. Oceanogr. 42 (Part 2 (5)), 1105–1118.
- Virnstein, R.W., Carbonara, P.A., 1985. Seasonal abundance and distribution of drift algae and seagrasses in the mid-Indian River Lagoon, Florida. Aquat. Bot. 23 (1), 67–82.

- Vitousek, P.M., Aber, J.D., Howarth, R.W., Likens, G.E., Matson, P.A., Schindler, D.W., Schlesinger, W.H., Tilman, D.G., 1997. Technical report: human alteration of the global nitrogen cycle: sources and consequences. Ecol. Appl. 7, 737–750.
- Wanielista, M.P., Yousef, Y.A., 1993. Stormwater Management. John Wiley & Sons Inc., New York, N.Y. 579 pp.
- Weiskel, P.K., Howes, B.L., 1992. Differential transport of sewage-driven nitrogen and phosphorus through a coastal watershed. Environ. Sci. Technol. 26 (2), 352–360.
- Wines, M., 2013. Deaths of manatees, dolphins and pelicans point to estuary at risk. The New York Times, In: http://www.nytimes.com/2013/08/08/us/

deaths-of-manatees-dolphins-and-pelicans-point-to-estuary-at-risk.html?. ref=manatees&\_r=0.

- Woodward-Clyde, 1994. Final Report, Historical imagery inventory and seagrass assessment Indian River Lagoon. Prepared for Indian River Lagoon National Estuary Program, Melbourne, FL. Project Number 92F274C 114 pp.
- Yarbro, L.A., Carlson Jr., P.R., 2013. Seagrass integrated mapping and monitoring program (SIMM): Mapping and monitoring report no 1. Fish and Wildlife Research Institute Technical Report TR-17 126 pp.
- Yentsch, C.S., Lapointe, B.E., Poulton, N., Phinney, D.A., 2008. Anatomy of a red tide bloom off the southwest coast of Florida. Harmful Algae 7 (6), 817–826.